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Human Brain Banking as a Convergence Platform of Neuroscience and Neuropsychiatric Research

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ABSTRACT

Neuropsychiatric disorders affect hundreds of millions of people and their families worldwide. Many studies have used human postmortem brain samples to decipher the molecular framework of these diseases. These studies uncovered brain-specific genetic and epigenetic patterns using highthroughput sequencing techniques. Therefore, determining the best way to collect human postmortem brain samples, analysing such a large amount of sequencing data, and interpreting these results are critical to advancing the field of neuropsychiatric sciences. By collecting postmortem/biopsied neural tissues and information about the diseases and life of donors, human brain banks support the observation and research of human brain sciences. Furthermore, the construction of large-scale brain banks has promoted the exploration of human brain morphology and function, development and ageing, as well as the mechanism of many neuropsychiatric diseases, which progressively reveal the normal mechanism of human brain activities and lead the direction of the prevention and treatment of neurological diseases. This article introduces the significance of human brain tissue bank construction and the current situation of the human brain tissue bank worldwide, as well as an overview of neurology or neuroscience advanced by using human brain samples.

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Human brain banks around the world

The research focused on human brains is essential to increase our understanding of neuropsychiatric disorders, not only in terms of disease pathogenesis but also to translate research on molecular mechanisms in disease models to novel treatments for human beings (Samarasekera et al., 2013). To meet researchers' demands, human brain banks have been established worldwide, and networks have been set up both in developing and developed countries. These networks have enabled a more focused approach to the study of neuropsychiatric disorders, with individual banks specialising in specific disorders and donor programs in clinical trials or population-based cohorts (Samarasekera et al., 2013).

The Alzforum (<u>https://www.alzforum.org</u>) is a nonprofit that provides information regarding human brain banks that have been constructed worldwide (<u>https://www.alzforum.org/brain-banks</u>). A total of

144 human brain banks were included: 81 brain banks in the United States, 16 brain banks in the United Kingdom, 10 brain banks in Australia, 7 brain banks in Germany, 4 brain banks in France, 3 brain banks in Canada, 3 brain banks in the Republic of Korea, 3 brain banks in Spain, 2 brain banks in Austria, 2 brain banks in Finland, 2 brain banks in Austria, 2 brain banks in Finland, 2 brain banks in Italy, 2 brain banks in Mexico, 1 brain bank in China, 1 brain bank in Greece, 1 brain bank in Hungary, 1 brain bank in India, 1 brain bank in Japan, 1 brain bank in the Netherlands, 1 brain bank in New Zealand, 1 brain bank in South Africa, and 1 brain bank in Sweden (Table 1). It is worth noting that the United States, the European Union, and Australia have established the brain bank consortium (Ma et al., 2019).

For example, the National Institutes of Health (NIH) established a specialised agency, the NeuroBioBank (NBB), to support and coordinate the National Brain Tissue Bank and to serve as a central point of access to the world-class collections of its six biorepositories: University of Miami Brain Endowment Bank, University of Maryland Brain and Tissue Bank, Harvard Brain Tissue Resource Center, the Human Brain and Spinal Fluid Resource Center, Mt. Sinai Brain Bank, and Brain Tissue Donation Program at the University of Pittsburgh (https://neurobiobank. nih.gov/about/#network). In addition, the BNE (BrainNet Europe) Consortium consists of nineteen European Brain Banks distributed in the United Kingdom, France, Germany, Spain, Italy, Denmark, Greece, Austria, Finland, Iceland, Sweden, Hungary, and other countries (Ma et al., 2019). The BNE Consortium was funded by the European Commission's Sixth Framework Program for the Life Sciences Project (LSHM-CT-2004-503039) to provide high-quality human brain samples, carry out quantitative research on human brain nucleic acids, proteins, and neurochemicals, and explore the normal working principles and disease mechanisms of the human brain (Ma et al., 2019). The BNE Consortium required members to adopt a unified and standardised procedure for brain sampling, preservation, and essential pathological detection and promoted the sharing of brain tissue resources to support collaborative neuroscience research in Europe and the rest of the world (Ma et al., 2019). In addition, the UK Brain Banks Network established a coordinated national network of UK brain tissue resources (banks) for researchers to use, including Cambridge Brain Bank, Edinburgh

Brain Banks, London Neurodegenerative Diseases Brain Bank, Manchester Brain Bank, Multiple Sclerosis and Parkinson's Tissue Bank, Newcastle Brain Tissue Resource, Oxford Brain Bank, Queen Square Brain Bank for Neuropsychiatric Disorders, Sheffield Brain Tissue Bank (SBTB), and South West Dementia Brain Bank (https:// brainbanknetwork.ac.uk/BrainBank/BrainBanks).

Development of human brain banks in China

Unlike developed countries, China's human brain bank has remained preliminary over decades (Ma et al., 2019). However, joint efforts since 2012 have substantially promoted this critical frontier of brain bank construction in China to support high-quality research into major neuropsychiatric, psychiatric, and developmental brain diseases affecting the Chinese people (Ma et al., 2019). First, the Standardised Operational Protocol for brain banking in China was published in 2019 after extensive collaboration among domestic and international experts over several years of continuous effort (Ma et al., 2019; Oiu et al., 2019). Second, three specific human brain banking conferences have been held to strengthen the scholarly consensus and coordinate multiinstitutional efforts on brain bank construction in China (Ma et al., 2019). In 2014, the first international workshop focused on brain bank construction was held in Changsha and Beijing. Professor Shumin Duan proposed establishing multiple banking centers and developing shared protocols and databases accessible to all researchers (Ma et al., 2019). Following this initiative, a second workshop was held in Beijing at Peking Union Medical College in 2016, and then a third workshop was held in Hangzhou at Zhejiang University in 2018 (Ma et al., 2019). All these meetings consisted of thematic presentations on topics covering the role of human brain banking in basic and translational neuroscience, the status of brain banking across the world, and current research findings in the areas of neuropsychiatric, and neurodevelopmental brain diseases (Ma et al., 2019). To date, China has already been establishing a nationwide brain bank alliance, the China Brain Bank Consortium. led by the National Human Brain Bank for Development and Function, Institute of Basic Medical Sciences Chinese Academy of Medical, Sciences Peking Union Medical College and National Health and Disease Human Brain Tissue

AMERICA							
Brain Banks	Location	Website	Details	Samples			
NIH-funded	United States	https://neurobiobank.	An open-access network of six established	N/A			
NeuroBioBank		<u>nih.gov/</u>	brain banks in the United States that stores				
			samples covering neurological, neuropsychi-				
			atric, and neurodevelopmental diseases and				
			disorders with good clinical information and				
			neuropathology, as well as normal controls.				
Mayo Clinic	Rochester, MN,	https://www.mayo.edu/	Collected over 3,000 cases of Alzheimer's	> 9,000			
Brain Bank	United States	research/departments-	disease, 2,000 cases with Lewy body	specimens			
		divisions/department-	disease, nearly 2,000 cases of tauopathies,				
		neuroscience-florida/	and a growing collection of frontotemporal				
		brain-banks/mayo-	dementia and ALS cases with well clinical				
		<u>clinic-brain-bank</u>	information and neuropathology, as well as				
			normal controls.				
Harvard	Belmont, MA,	https://hbtrc.mclean.	Collected over 9,000 donated brains with	> 9,000			
Brain	United States	harvard.edu/	well clinical information and neuropathology	specimens			
Tissue			information, as well as normal controls, and				
Resource			distributed over a hundred thousand samples,				
Center			both United States nationally and globally.				
Human Brain	Rockville, MD,	https://www.nimh.nih.	Collected human brain tissue and blood	> 1,000			
Collection Core	United States	gov/research/research-	samples from deceased individuals	specimens			
(HBCC)		conducted-at-nimh/re-	diagnosed with mental illnesses such as				
		search-areas/research-	schizophrenia, depression, bipolar disorder,				
		support-services/hbcc	substance abuse disorder, and those who				
			died by suicide, as well as from individuals				
	~		without a history of mental illness				
UCSF Neuro-	San Francisco,	https://memory.ucsf.	Established in 2008 and serves as a reposi-	N/A			
degenerative	CA, United	edu/research-trials/	tory for nervous system tissue donated for				
Disease Brain	States	professional/neuro-	research purposes. NDBB is committed to				
Bank (NDBB)		degenerative-disease-	advancing the understanding of neurodegen-				
		brain-bank	erative disease by (1) performing compre-				
			hensive neuropathological characterisation				
			of participants who participated in clinical				
			research during life and (2) providing tissue				
			to leading investigators worldwide.				
EUROPE							
Brain Banks	Location	Website		Samples			
BrainNet	Munich,	https://cordis.europa.	A network of 19 established brain banks	N/A			
Europe	Germany	eu/project/1d/503039	across Europe that stores samples covering				
			neuropsychiatric disorders with good				
			clinical information and neuropathology				
			information, as well as normal controls.	> 1.000			
Netherlands	Amsterdam,	<u>nttps://www.brainbank.</u>	An open-access brain bank that collected	> 4,000			
Brain Bank	Netherlands	ni/brain-tissue/avaii-	numan brain tissue of donors with a variety	specimens			
		<u>aomty/</u>	of neurological and psychiatric disorders				
D · C	T 1	1	but also of nondiseased donors.				
Brains for	London,	https://www.kci.ac.uk/	Established in 2007 to promote brain	IN/A			
Dementia	Vined	research/brains-lor-	donation and develop a network of brain				
Research,	Kingdom	dementia-research	dissue banks to facilitate research into				
for A so Delated			dementia.				
Diseases							
LIV Droin	Madiaal Da	https://hpsiphenlagt	A appreciated notional returned a CIUV has been				
UK Brain	wiedical Ke-	<u>mups://orainbanknet-</u>	A coordinated national network of UK brain				
Danks Notwork	Bristol Unite 1	work.ac.uk/BrainBank/	ussue resources (danks).				
INCLWOIK	Kingdom						
	Kinguom						

 Table 1.
 Human brain banks around the world.

ASIA							
Brain Banks	Location	Website	Details	Samples			
National Human	Beijing, China	http://anatomy.sbm.	As national resource platform, support and	> 500			
Brain Bank for		pumc.edu.cn/brain-	lead China's brain science research, and	specimens			
Development		<u>bank/</u>	make positive contributions to maintaining				
and Function			brain health and defeating brain diseases.				
National Human	Hangzhou,	http://zjubrainbank.zju.	Collected and stored more than 300 human	> 300			
Brain Bank	Zhejiang, China	edu.cn/index	whole brain samples, covering common neuro-	specimens			
Disease			hybrid brain complex without brain disease				
Central South	Changsha	https://www.csu.edu	A member of the China Brain Bank	N/A			
University	Hunan China	nups.//xysin.csu.cuu.	Consortium	IN/A			
Brain Bank	Trunan, China		consortium.				
Fudan Univer-	Shanghai	https://shmc_fudan	A member of the China Brain Bank	N/A			
sity Brain Bank	China	edu.cn/	Consortium.				
Tiantan Hospital	Beijing, China	https://bjtth.org/	A member of the China Brain Bank	N/A			
Capital Medical			Consortium.				
University Brain							
Bank							
Hebei Medi-	Shijiazhuang,	https://www.hebmu.	A member of the China Brain Bank	N/A			
cal University	Hebei, China	edu.cn/	Consortium.				
Brain Bank							
Anhui Medi-	Hefei, Anhui,	https://www.ahmu.	A member of the China Brain Bank	N/A			
cal University	China	edu.cn/	Consortium.				
Brain Bank							
Guiyang Medi-	Guiyang,	https://gmc.edu.cn/	A member of the China Brain Bank	N/A			
cal University	Guizhou, China		Consortium.				
Brain Bank		1					
Brain Bank for	Tokyo, Japan	https://www.tmig.or.jp/	For aging and dementia and construction of	N/A			
Aging Research	V ann atalaa	eresearch/a23.ntml	Genatric Neuroscience Data Base.	> 715			
Human Brain	Karnataka,	hank in/brain bank/	A resource organisation to procure precious	> /15			
Dalik	Illula	<u>Ualik.iii/Ulalii-Ualik/</u>	research	specificits			
		AUSTRALI	A/OCEANIA	4			
Brain Banks	Location	Website	Details	Samples			
Victorian Brain	VIC. Australia	https://florey.edu.au/	Supports research into the study of brain	N/A			
Bank		science-research/scien-	disorders by providing tissue to researchers.				
		tific-services-facilities/					
		victorian-brain-bank					
MS Research	NSW, Australia	https://msbrainbank.	Over 2260 people with MS have registered	> 80			
Australia Brain		org.au/progress/	their interest and more than 930 have con-	specimens			
Bank			sented to their tissue.				
NSW Brain	Australia	http://www.nswbrain-		N/A			
Bank Network		banknetwork.org.au/					
Schizophre-	NSW, Australia	https://www.neura.edu.	Collects and links genetic, clinical, neuro-	> 1,000			
nia Research		au/discovery-portal/	psychological and brain imaging infor-	specimens			
Institute		<u>asrb/</u>	mation from over 1,000 individuals with				
Conder and Darain	NCW Assetselie	1	schizophrenia and healthy controls.	200			
Sydney Brain	NSW, Australia	<u>nups://www.neura.edu.</u>	Hold brain ussue from over /00 donors with	> /00			
Dalik		<u>au/syuncyuraniuank/</u>	disease Lewy body disease motor neuron	specificits			
			disease, Huntington's disease, frontotemporal				
			lobar degeneration multiple system atrophy				
Neurological	AUK New	https://neurological	Received more than 700 brains and brain	> 700			
Foundation of	Zealand	org.nz/what-we-do/	tissue samples from over 300 donors with	specimens			
New Zealand		neurological-founda-	neurosurgical operations.				
Human Brain		tion-human-brain-bank/					
Bank							
		AFF	RICA				
Brain Banks	Location	Website	Details	Samples			
Tygerberg	Cape Town,	http://www.trauma.org/		N/A			
Hospital	South Africa	index.php/resources/					
1		elective/352/		1			

Resource Center, Zhejiang University, and other members, including Shanghai Medical College Fudan University, Xiangya School of Medicine of Central South University, Beijing Tian Tan Hospital Capital Medical University, Hebei Medical University and so on. The mission of the China Brain Bank Consortium is to advance basic and translational human brain research by providing donors clinical biometrics, brain/tissue/ fluid samples, and primary neuropathological documentation (Ma et al., 2019). The China Brain Bank Consortium also aims to serve as a knowledge-integration and broadcasting organisation for public education and policy decision-making to improve neuropsychiatric and neurodevelopmental brain disease research and, ultimately, the brain health and life of the Chinese people (Ma et al., 2019).

Human brain tissue as a cornerstone of neuroscience

To help researchers review the history of the development of research and make further contributions to future research in the brain field, a total of 7172 articles were retrieved from the Web of Science database using the following strategy: "Topic = postmortem brain", and publication years were between 2012 and 2021. Fig. 1A shows the Number of articles published from 2012 - 2021. The total citations of articles published from 2012 - 2021 are shown in Fig. 1B. The distribution of research focus of 7172 articles between 2012 and 2021 were unevenly distributed, mainly focused on neuroscience, neurology, biochemistry, molecular biology, psychiatry, and behavioural sciences (Fig. 1C). The use of



Figure 1. Analysis of articles citing postmortem brain between 2012 and 2021. A: Number of articles published from 2012 - 2021. B: Citation analysis of articles published from 2012 - 2021. C: Research foci of articles citing postmortem brain in 2012 - 2021.

postmortem human brain materials has given insights into brain differences in health-related to age, sex, sexual orientation and human brain diseases, from neurodegenerative diseases and neuroendocrine disorders to psychiatric disorders, especially Alzheimer's disease, Parkinson's disease, schizophrenia and bipolar disorder.

Utilising human brain tissue sections from dementia patients, argyrophilic senile plaques and neurofibrillary tangles (NFTs) of the cerebral cortex were found by Alois Alzheimer in 1906 (Alzheimer, et al., 1995; Morris & Salmon, 2007). Dementia, which is characterised by the deposition of argyrophilic senile plaques and NFTs in the brain, was named Alzheimer's disease (AD) (Montine et al., 2012). A spate of molecular neurobiology research identified inheritable mutations of the APP gene directly responsible for familial AD (Campion et al., 1999). In singletransgenic or double-transgenic animal models (Tg animal) of APP mutations of familial AD (Borchelt et al., 1997; Carlson et al., 1997; Games et al., 1995), the histopathological progression of cerebral amyloid- β (A β) deposition was reproduced, and the synaptic damage, neuronal loss and memory impairment of the AD patients can again be found in the animal models of AD, but NFTs, the major neuropathological features of AD brain, cannot be reproduced in the Tg animals (Pang et al., 2022; Sanchez-Varo et al., 2022; Sasaguri et al., 2017). The information that was obtained from the collected human brain tissues was indispensable for uncovering the mechanisms of neuropsychiatric disorders. Without well-preserved human brain tissues that were a standardised collection and had perfect clinical information, understanding the agents of neuropsychiatric disorders would have been impossible.

In recent years, more knowledge of neuropsychiatric disorders has been obtained from cultured cells, brain organoids and Tg animals with the progress and application of new technologies. Over the years, Tg animal models and cell research have overshadowed human tissues, especially brain or other nerve tissues (Duyckaerts et al., 2008; Webster et al., 2014). Animal models and cellular experiments have many advantages, such as ease of manipulation. convenience for longitudinal studies, more observations performed in living animals, and gene knockout and transgene experiments. Cellular culture and animal models cannot always mimic every aspect of human disease, especially mental and neuropsychiatric disorders (Duyckaerts et al., 2008). For example, monogenic and biogenic APP-based AD mouse models involving mutant APP or mutant APP and presenilin (PS) 1 or presenilin 2 gene expression model the typical amyloid- β (A β) accumulation of familial and sporadic AD in humans, but neuropathological changes in NFTs have not been found (Carlson et al., 1997; Duyckaerts et al., 2008; Sanchez-Varo et al., 2022; Sasaguri et al., 2017; Webster et al., 2014). Although current Tg-mice of AD models reproduce the $A\beta$ plaques that were described in human AD brains to some extent, the morphological variety and diversity of A β plaques are wider in human samples. According to current studies, A β , tau, and ApoE are key molecules in AD (Borchelt et al., 1997; Borchelt et al., 1996; Fryer et al., 2005; Funato et al., 1998; Gessel et al., 2012; Lewis et al., 2001; Lewis et al., 2000; Maruyama et al., 2013). Nevertheless, AD research has benefited from various transgenic mice, such as the function, metabolism and behaviour of critical molecules relevant to AD. For example, Nav1.1 hypofunction has been observed in mouse models of AD (Corbett et al., 2013; Hamm et al., 2017), and the effect of A β amyloid on Nav1.1 phenotypic consequences should be reviewed and validated in future studies. However, the genes and proteins of A β , tau, and ApoE differ between rodents and humans in their sequences, pathogenicity or number of isoforms expressed.

PD is a neurodegenerative disorder that affects approximately 1.5% of the global population over 65 years of age, the hallmark feature of which is degeneration of the dopamine (DA) neurons in the substantia nigra pars compacta (SNc) and the consequent striatal DA deficiency (Brichta et al., 2013; Nussbaum & Polymeropoulos, 1997; Pringsheim et al., 2014; Rodriguez-Oroz et al., 2009). From human patients with familial PD, researchers found the first PD-linked gene SNCA, code synuclein alpha (α -syn), which is a member of the synuclein family and the main component of Lewy bodies (Chan et al., 1998; Farrer et al., 1998; Munoz et al., 1997; Nussbaum & Polymeropoulos, 1997; Zareparsi et al., 1998). Although there is currently no cure for PD. multiple mutations of PD-related genes were found, such as three missense mutations (encoding

the substitutions A30P, A53T, and E46K) of the SNCA gene of familial PD (Chan et al., 1998; Farrer et al., 1998; Munoz et al., 1997; Zareparsi et al., 1998), mutations of the PARKIN gene of familial PD and some young-onset sporadic PD patients (Abbas et al., 1999; Hattori et al., 1998: Kitada et al., 1998: Lucking et al., 2000). mutations (G2019S and R1441C) of the LRRK2 (Leucine rich repeat kinase 2) gene of the lateonset autosomal dominant inherited form of PD (Di Fonzo et al., 2005; Funayama et al., 2005; Gilks et al., 2005; Nichols et al., 2005; Zimprich et al., 2004), and mutations of the PINK1 (PTEN Induced Putative Kinase 1) gene or DJ-1 gene in early-onset PD (Abou-Sleiman et al., Wood, 2004; Bonifati et al., 2005; Deng et al., 2005; Healy et al., 2004; C. Klein et al., 2005; McInerney-Leo et al., 2005; Rogaeva et al., 2004; Valente et al., 2004). Genetic animal models of PD can better understand the mechanisms underlying the genetic forms of PD (Beal, 2001; Blesa et al., 2012; Blesa & Przedborski, 2014; Chesselet & Richter, 2011; Metzger & Emborg, 2019). However, the neuropathology and behavioural phenotypes of PD animal models are often quite different from the human condition (Hinkle et al., 2012; Rathke-Hartlieb et al., 2001; Sanchez et al., 2014). Thus, transgenic models of animals with mutated genes observed in familial PD patients are insufficient to reproduce the final neuropathological features of PD. In most populations, the known pathogenic mutations of SNCA, Parkin, PINK1, and DJ1 are infrequent and can only explain a minor part of all PD cases (Hinkle et al., 2012; Rathke-Hartlieb et al., 2001; Sanchez et al., 2014).

Similar to AD and PD, it seems that many neuropsychiatric disorders, such as Parkinson's disease (PD), schizophrenia, and bipolar disorders, may be specific to humans, have no real equivalent, or are unknown in animals. Neuropsychiatric disorders often have subjective symptoms and a variety of neuropathological changes in the brain, which are unique challenges for animal models of CNS disorders. Even though the technology of cell and tissue culture from postmortem tissue was discovered more than 40 years ago (Macpherson et al., 1985), it has rapidly evolved in recent decades (Dimos et al., 2008; Ebert et al., 2009; Israel et al., 2012; Marchetto et al., 2010; X. Oian et al., 2016; Soldner et al., 2009; Takahashi et al., 2007). The discovery of induced pluripotent stem cells (iPSCs) from

into various cell types, such as neurons, has allowed scalable testing of neurodegenerationassociated genes and therapeutic agents entirely in human systems (Ebert et al., 2009; Israel et al., 2012; Marchetto et al., 2010; Qian et al., 2016; Soldner et al., 2009). Neurons that are currently created from iPSCs have been applied to study the underlying mechanisms of spinal muscular atrophy, amyotrophic lateral sclerosis, AD and schizophrenia (Choi et al., 2014; Egawa et al., 2012; Israel et al., 2012; Lee et al., 2009; Marchetto et al., 2010; Mariani et al., 2015; Miller et al., 2013; Moore et al., 2002; X. Qian et al., 2016; Soldner et al., 2009; Zhang et al., 2013). With the rise and use of new technologies such as brain organoid culture, cell and tissue culture from autopsy materials has occurred for psychiatric research. Cultured brain organoids from human postmortem tissue have been used to study complex neuropsychiatric diseases such as AD, frontotemporal dementia (FTD), PD, motor neuron disease and schizophrenia with neurodegeneration (Di Lullo & Kriegstein, 2017; Eiraku et al., 2011; Eiraku et al., 2008; Jo et al., 2016; Kawada et al., 2017; Lancaster et al., 2013; Mellios et al., 2018; Monzel et al., 2017; Nakano et al., 2012; Pasca et al., 2015; X. Y. Oian et al., 2018: Stachowiak et al., 2017). Brain organoid culturing offers a promising avenue to study the underlying genetic mechanism of neuropsychiatric disorders. Brain organoids or cerebral organoid cultures, which are threedimensional (3D) cultures, have more precise neural layering and identity and allow tuning and generation of regional brain tissue structures such as cerebral, hypothalamus, midbrain, cerebellar, and hippocampal-specific organoids that enable elucidation of regional pathological mechanisms specific to more complex neuropsychiatric disorders (Hor et al., 2018; Kawada et al., 2017; Muguruma et al., 2010; Sakaguchi et al., 2015). iPSCs or brain organoids obtained by postmortem tissue can offer a unique advantage in terms of gene expression profiles and epigenetic properties of neurons. The gene expression profiles and epigenetic properties of human postmortem brain tissue allow further validation of the accuracy of iPSC-derived neurons or brain organoids. Furthermore, iPSCs or brain organoids from postmortem tissue also create methods to study the postmortem abnormalities observed in vitro and in vivo.

Brain bank advancing neuropsychiatric disease studies

Collecting human brains has a pivotal role in understanding the biological basis of the brain and neuropsychiatric diseases. The knowledge of the brain essence and nervous system disease is inseparable from the direct study of human brain material. The human brain bank collects postmortem or biopsied human nerve tissue samples of donors, including antemortem illness and life information, and shares samples and attendant demographic and clinical data with qualified researchers worldwide for research on neuroscience and neuropsychiatric diseases (Deep-Soboslay et al., 2011; Rademaker et al., 2018; Neshika Samarasekera et al., 2013; L. Wang et al., 2019). The human brain bank s a scientific platform to support the direct observation and research of health and disease brain tissue. Transcriptomics, proteomics, and lipidomics are applied to study the neuropathology mechanism of neuropsychiatric diseases and to search for new clinical therapeutic targets. The human brain, which is a total export of molecular omics (proteomics, transcriptomics, and genomics), histocytology, neuropathology, imageology, and neuropsychiatry in clinical neuroscience, will provide conditions and opportunities for revealing the workings of the brain and the mechanisms of neuropsychiatric disease. Animal models of neuropsychiatric disorders or cell and tissue culture will confirm new findings from human brain tissues. At the same time, data from animals or cells and tissue must also be validated on human brain tissue. In addition, with the traditional use of human brain samples for histopathological studies, many new approaches and methods are being developed for basic and translational neuroscience research by testing brain samples.

High levels of the fibrillogenic A β 1-42 peptides were found to be preferentially deposited in the brains of AD patients (Borchelt et al., 1996; Iwatsubo et al., 1994). Since the 1990s, mutations in the APP, PSEN1, and PSEN2 genes are responsible for A β deposition in the earlyonset autosomal dominant familial forms of AD (Borchelt et al., 1996; Citron et al., 1992; Ertekin-Taner, 2010; St George-Hyslop et al., 1987). Genome-wide association studies (GWASs) confirmed APOE as a genetic risk factor for lateonset AD (LOAD). In addition to APOE, the Alzheimer Disease Genetics Consortium (ADGC) and other researchers further reported nine novel LOAD susceptibility loci, including CR1, CLU, PICALM, BIN1, EPHA1, MS4A, CD33, CD2AP and ABCA7, by using genome-wide association studies (GWASs) (Hollingworth et al., 2011; Naj et al., 2011). These novel disease loci of AD identified in the recent large disease GWAS may uncover the pathophysiology of LOAD. Combined brain gene expression endophenotype and disease GWAS results revealed that the LOAD SNPs at the CLU and MS4A loci influence brain gene expression of CLU and MS4A4A genes, and additional variants within the arbitrary 100 kb cis-region also influence brain expression of CLU and ABCA7 (Allen et al., 2012). Brain expression changes in CLU, MS4A4A and ABCA7 may partly explain the LOAD risk association at these loci (Allen et al., 2012). By measuring the expression levels of transcripts in brain samples from the cerebellum and temporal cortex of autopsied subjects with AD, a combined assessment of expression and disease GWAS reported that 2,980 cerebellar cisSNP/transcript level associations (2,596 unique cisSNPs) were significant in both AD and non-AD (Zou et al., 2012). A recent assessment of postmortem human brain tissue revealed that upregulation of histone acetyltransferases H3K27ac and H3K9ac in the lateral temporal lobe was linked to disease pathways in AD by dysregulating transcriptionand chromatin-gene feedback loops with an integrated multiomics approach (Nativio et al., 2020). An epigenome-wide association study with the histone 3 lysine 9 acetylation (H3K9ac) mark revealed that tau protein burden broadly affects histone acetylation in the aged and AD human brain (H. U. Klein et al., 2019). The human microglial transcriptome and genetic analysis of postmortem brain samples obtained from the Netherlands Brain Bank (NBB) and the Neuropathology Brain Bank showed associations with microglial expression of USP6NL for Alzheimer's disease and P2RY12 for Parkinson's disease (Lopes et al., 2022). High-throughput integrative transcriptome expression analyses of postmortem brain tissue indicated that the complement pathway plays a crucial role in the protective effect of APOE $\varepsilon 2$ on AD (Panitch et al., 2021). STMN2, which encodes a stathmin family protein, is a key regulator functionally connected to known PD risk genes (Wang et al., 2019), and both bipolar disorder and schizophrenia revealed convergent

changes in cortical astrocytes and fast-spiking parvalbumin interneurons (Toker et al., 2018). Transcriptome data from 1,865 brain samples showed that the edited site in the 3'-UTR of SYT11 may influence SYT11 protein levels in aged and AD human brain tissue (Ma et al., 2021). Synaptic transmission deficits of AD were first discovered in the 1970s when reductions in cholinergic markers were observed in the brains of patients diagnosed with AD (Bowen, Smith, White, & Davison, 1976). Several studies have revealed a series of synaptic protein changes in various types of dementia by using the postmortem human brain (Bereczki et al., 2018; Bereczki et al., 2016; Francis et al., 2019; Vallortigara et al., 2014; Vallortigara et al., 2016; Whitfield et al., 2014). Structural variants (SVs) that are essential sources of genetic diversity have been linked to many diseases, such as AD. Using 1,760 whole genomes from aged and AD individuals, 170,996 SVs were discovered. Quantitative trait locus (SVxOTL) analyses revealed that more than 3,200 SVs were associated with at least one molecular phenotype in postmortem brain tissues, such as histone modifications, gene expression, splicing and protein abundance (Vialle et al., 2022).

Conclusions and future directions

In the past ten years, there have been significant developments in regulatory frameworks governing brain banking in China and the world, especially methods for optimising tissue collection and the range of neuropathological techniques available for analysing brain tissue. These novel findings in genomic, transcriptomic, proteomics and epigenomic studies of the brain highlight the potential of stored brain tissue to advance our understanding of significant scientific and clinical questions (Allen et al., 2012; Colantuoni et al., 2011; Ertekin-Taner, 2010; Hollingworth et al., 2011; H. U. Klein et al., 2019; Lopes et al., 2022; Naj et al., 2011; Nativio et al., 2020; Panitch et al., 2021; Qi et al., 2022; Toker et al., 2018; Vialle et al., 2022; Q. Wang et al., 2019; Xiong, Ge, & Ma, 2019; Zou et al., 2012). With the application of new technologies such as single-cell RNA sequencing, the application potential of human brain tissue stored in brain banks will be further enhanced. It is anticipated that basic translational studies of human brain tissue will be promoted with further scientific advances, specifically in animal models of neuropsychiatric diseases or cell culture models.

Brain tissue from spontaneous donations is predominantly from end-stage disease, and information on the disease at earlier stages is uncommon. Control tissues are required to support neuroscience research. Although more than enough people are willing to donate their brains to scientific research, recruitment of control cases is still problematic and restricted. Very few national programs or organisations focus on obtaining healthy control brains. Participants in longitudinal cohorts should be encouraged to donate. The advantages of donations from participants in this cohort are the wide range of clinical information available (including MRI data, genetic data, biomarkers and cognitive function), detailed information on lifestyle and comorbidities, and the potential to obtain samples from patients who are likely to die early in the disease.

Fetal and infant brain collection is an important and challenging direction for brain collection. At most human brain banks, donated human brains come from older people and are suitable for research into neurodegenerative diseases. Nevertheless, for neurodevelopmental disorders such as autism, SCZ, and intellectual disabilities, fetal and infant brain samples are essential for researching the causes of the disorders. Only a few banks have prenatal samples, and the sample size is comparatively limited.

Many nervous system diseases are systemic, and researchers are increasingly interested in obtaining tissue samples other than those of the brain. Therefore, samples from the appendix, colon and intestines may yield new insights. The immune system plays a role in diseases such as multiple sclerosis and AD and other neurodegenerative diseases, so the availability of blood, immune cells or immune organs can be significant.

The ultimate measure of the success of a professional brain bank is the outcome and quality of the research conducted on preserved brain samples. Progress is likely to be driven by collaborative networks that promote brain banks, which will increase sample sizes for research on rare neuropsychiatric diseases, develop generally applicable standards for postmortem brain tissue research and share ideas to maximise the potential of storing brain tissue as a valuable research resource.

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Conflict of interest

There are no potential conflicts in this research's financial and material support.

Reference

- Abbas, N., Lucking, C. B., Ricard, S., Durr, A., Bonifati, V., De Michele, G., . . . European Consortium Genetic Susceptibility, P. (1999). A wide variety of mutations in the parkin gene are responsible for autosomal recessive parkinsonism in Europe. *Human Molecular Genetics*, 8(4), 567-574. doi: 10.1093/hmg/8.4.567
- Abou-Sleiman, P. M., Healy, D. G., & Wood, N. W. (2004). Causes of Parkinson's disease: genetics of DJ-1. *Cell and Tissue Research*, *318*(1), 185-188. doi:10.1007/s00441-004-0922-6
- Allen, M., Zou, F., Chai, H. S., Younkin, C. S., Crook, J., Pankratz, V. S., . . . Woltjer, R. L. (2012). Novel late-onset Alzheimer disease loci variants associate with brain gene expression. *Neurology*, 79(3), 221-228. doi:10.1212/WNL.0b013e3182605801
- Alzheimer, A., Stelzmann, R.A., Schnitzlein, H.N.,
 & Murtagh, F. R. (1995). An English translation of Alzheimer's 1907 paper, "Uber eine eigenartige Erkankung der Hirnrinde". *Clinical Anatomy*, 8(6), 429-431. doi: 10.1002/ca.980080612
- Beal, M. F. (2001). Experimental models of Parkinson's disease. *Nature Reviews Neuroscience*, 2(5), 325-332. doi:10.1038/35072550
- Bereczki, E., Branca, R. M., Francis, P. T., Pereira, J. B., Baek, J. H., Hortobágyi, T., . . . Aarsland, D. (2018). Synaptic markers of cognitive decline in neurodegenerative diseases: a proteomic approach. *Brain*, *141*(2), 582-595. <u>doi:10.1093/brain/ awx352</u>
- Bereczki, E., Francis, P.T., Howlett, D., Pereira, J.B., Höglund, K., Bogstedt, A., . . . Aarsland, D.

(2016). Synaptic proteins predict cognitive decline in Alzheimer's disease and Lewy body dementia. *Alzheimers & Dementia, 12*(11), 1149-1158. doi:10.1016/j.jalz. 2016.04.005

- Blesa, J., Phani, S., Jackson-Lewis, V., & Przedborski, S. (2012). Classic and New Animal Models of Parkinson's Disease. *Journal of Biomedicine and Biotechnology*. doi:10.1155/2012/845618
- Blesa, J., & Przedborski, S. (2014). Parkinson's disease: animal models and dopaminergic cell vulnerability. *Frontiers In Neuroanatomy*, 8, 155. <u>doi:10.3389/fnana.</u> 2014.00155
- Bonifati, V., Rohe, C. F., Breedveld, G. J., Fabrizio, E., De Mari, M., Tassorelli, C., . . . Italian Parkinson, N. (2005). Earlyonset parkinsonism associated with PINK1 mutations - Frequency, genotypes, and phenotypes. *Neurology*, 65(1), 87-95. doi:10.1212/01.wnl.0000167546.39375.82
- Borchelt, D. R., Ratovitski, T., van Lare, J., Lee, M. K., Gonzales, V., Jenkins, N. A., . . . Sisodia, S. S. (1997). Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins. *Neuron*, *19*(4), 939-945. <u>doi:10.1016/s0896-6273</u> (00)80974-5
- Borchelt, D. R., Thinakaran, G., Eckman, C. B., Lee, M. K., Davenport, F., Ratovitsky, T., . . . Younkin, S. G. (1996). Familial Alzheimer's disease-linked presenilin 1 variants elevate A beta 1-42/1-40 ratio in vitro and in vivo. *Neuron*, *17*(5), 1005-1013. doi:10.1016/s0896-6273(00)80230-5
- Borchelt, D. R., Thinakaran, G., Eckman, C. B., Lee, M. K., Davenport, F., Ratovitsky, T., . . . Sisodia, S. S. (1996). Familial Alzheimer's disease-linked presenilin 1 variants elevate Abeta1-42/1-40 ratio in vitro and in vivo. *Neuron*, *17*(5), 1005-1013. doi:10.1016/s0896-6273(00)80230-5
- Bowen, D. M., Smith, C. B., White, P., & Davison, A. N. (1976). Neurotransmitterrelated enzymes and indices of hypoxia in senile dementia and other abiotrophies. *Brain*, 99(3), 459-496. <u>doi:10.1093/</u> <u>brain/99.3.459</u>
- Brichta, L., Greengard, P., & Flajolet, M. (2013). Advances in the pharmacological treatment of Parkinson's disease:

targeting neurotransmitter systems. *Trends in Neurosciences, 36*(9), 543-554. doi:10.1016/j.tins.2013.06.003

- Campion, D., Dumanchin, C., Hannequin, D., Dubois, B., Belliard, S., Puel, M., . . . Frebourg, T. (1999). Early-onset autosomal dominant Alzheimer disease: prevalence, genetic heterogeneity, and mutation spectrum. *American journal* of human genetics, 65(3), 664-670. doi:10.1086/302553
- Carlson, G. A., Borchelt, D. R., Dake, A., Turner, S., Danielson, V., Coffin, J. D., . . . Hsiao, K. K. (1997).Geneticmodificationofthephenotypes produced by amyloid precursor protein overexpression in transgenic mice. *Human Molecular Genetics*, 6(11), 1951-1959. doi:10.1093/hmg/6.11.1951
- Chan, P., Tanner, C. M., Jiang, X., & Langston, J. W. (1998). Failure to find the alphasynuclein gene missense mutation (G(209) A) in 100 patients with younger onset Parkinson's disease. *Neurology*, 50(2), 513-514. doi:10.1212/wnl.50.2.513
- Chesselet, M.-F., & Richter, F. (2011). Modelling of Parkinson's disease in mice. *Lancet Neurology*, *10*(12), 1108-1118. doi:10.1016/s1474-4422(11)70227-7
- Choi, S. H., Kim, Y. H., Hebisch, M., Sliwinski, C., Lee, S., D'Avanzo, C., . . . Kim, D. Y. (2014). A three-dimensional human neural cell culture model of Alzheimer's disease. *Nature*, 515(7526), 274-U293. doi:10.1038/nature13800
- Citron, M., Oltersdorf, T., Haass, C., McConlogue, L., Hung, A. Y., Seubert, P., . . . Selkoe, D. J. (1992). Mutation of the betaamyloid precursor protein in familial Alzheimer's disease increases beta-protein production. *Nature*, *360*(6405), 672-674. doi:10.1038/360672a0
- Colantuoni, C., Lipska, B. K., Ye, T., Hyde, T. M., Tao, R., Leek, J. T., . . . Kleinman, J. E. (2011). Temporal dynamics and genetic control of transcription in the human prefrontal cortex. *Nature*, *478*(7370), 519-523. doi:10.1038/nature10524
- Corbett, B. F., Leiser, S. C., Ling, H.-P., Nagy, R., Breysse, N., Zhang, X., . . . Chin, J. (2013). Sodium Channel Cleavage Is Associated with Aberrant Neuronal Activity and Cognitive Deficits in a Mouse Model of Alzheimer's Disease. *Journal*

of Neuroscience, 33(16), 7020-7026. doi:10.1523/jneurosci.2325-12.2013

- Deep-Soboslay, A., Benes, F. M., Haroutunian, V., Ellis, J. K., Kleinman, J. E., & Hyde, T. M. (2011). Psychiatric Brain Banking: Three Perspectives on Current Trends and Future Directions. *Biological Psychiatry*, 69(2), 104-112. <u>doi:10.1016/j.</u> <u>biopsych.2010.05.025</u>
- Deng, H., Le, W. D., Zhang, X., Pan, T. H., & Jankovic, J. (2005). G309D and W437OPA PINK1 mutations in Caucasian Parkinson's disease patients. *Acta Neurologica Scandinavica*, *111*(6), 351-352. <u>doi:10.1111/j.1600-0404.2005.00383.x</u>
- Di Fonzo, A., Rohe, C. F., Ferreira, R. J., Chien, H. F., Vacca, L., Stocchi, F., . . . Italian Parkinson Genetics, N. (2005). A frequent LRRK2 gene mutation associated with autosomal dominant Parkinson's disease. *Lancet*, 365(9457), 412-415. Retrieved from ≤Go to ISI>:// WOS:000226610900028
- Di Lullo, E., & Kriegstein, A. R. (2017). The use of brain organoids to investigate neural development and disease. *Nature Reviews Neuroscience*, 18(10), 573-584. doi:10.1038/nrn.2017.107
- Dimos, J. T., Rodolfa, K. T., Niakan, K. K., Weisenthal, L. M., Mitsumoto, H., Chung, W., . . . Eggan, K. (2008). Induced pluripotent stem cells generated from patients with ALS can be differentiated into motor neurons. *Science*, 321(5893), 1218-1221. doi:10.1126/science.1158799
- Duyckaerts, C., Potier, M. C., & Delatour, B. (2008). Alzheimer disease models and human neuropathology: similarities and differences. *Acta Neuropathologica*, *115*(1), 5-38. doi:10.1007/s00401-007-0312-8
- Ebert, A. D., Yu, J., Rose, F. F., Jr., Mattis, V. B., Lorson, C. L., Thomson, J. A., & Svendsen, C. N. (2009). Induced pluripotent stem cells from a spinal muscular atrophy patient. *Nature*, 457(7227), 277-280. doi:10.1038/nature07677
- Egawa, N., Kitaoka, S., Tsukita, K., Naitoh, M., Takahashi, K., Yamamoto, T., . . . Inoue, H. (2012). Drug Screening for ALS Using Patient-Specific Induced Pluripotent Stem Cells. *Science Translational Medicine*, 4(145). doi:10.1126/scitranslmed.3004052

- Eiraku, M., Takata, N., Ishibashi, H., Kawada, M., Sakakura, E., Okuda, S., . . . Sasai, Y. (2011). Self-organizing optic-cup morphogenesis in three-dimensional culture. *Nature*, 472(7341), 51-U73. doi:10.1038/nature09941
- Eiraku, M., Watanabe, K., Matsuo-Takasaki, M., Kawada, M., Yonemura, S., Matsumura, M., . . . Sasail, Y. (2008). Self-Organized Formation of Polarized Cortical Tissues from ESCs and Its Active Manipulation by Extrinsic Signals. *Cell Stem Cell*, 3(5), 519-532. doi:10.1016/j.stem.2008.09.002
- Ertekin-Taner, N. (2010). Genetics of Alzheimer disease in the pre- and post-GWAS era. *Alzheimer's Research & Therapy, 2*(1), 3-3. doi:10.1186/alzrt26
- Farrer, M., Wavrant-De Vrieze, F., Crook, R., Boles, L., Perez-Tur, J., Hardy, J., . . . Lynch, T. (1998). Low frequency of alpha-synuclein mutations in familial Parkinson's disease. *Annals of Neurology*, 43(3), 394-397. doi:10.1002/ana.410430320
- Francis, P. T., Hayes, G. M., Costello, H., & Whitfield, D. R. (2019). Brains for Dementia Research: The Importance of Cohorts in Brain Banking. *Neuroscience Bulletin*, 35(2), 289-294. doi:10.1007/ s12264-018-0327-2
- Fryer, J. D., Simmons, K., Parsadanian, M., Bales, K. R., Paul, S. M., Sullivan, P. M., & Holtzman, D. M. (2005). Human apolipoprotein E4 alters the amyloid-beta 40:42 ratio and promotes the formation of cerebral amyloid angiopathy in an amyloid precursor protein transgenic model. *Journal of Neuroscience*, 25(11), 2803-2810. doi:10.1523/jneurosci.5170-04.2005
- Funato, H., Yoshimura, M., Kusui, K., Tamaoka, A., Ishikawa, K., Ohkoshi, N., . . . Ihara, Y. (1998). Quantitation of amyloid betaprotein (A beta) in the cortex during aging and in Alzheimer's disease. *American Journal of Pathology*, 152(6), 1633-1640.
- Funayama, M., Hasegawa, K., Ohta, E., Kawashima, N., Komiyama, M., Kowa, H., . . Obata, F. (2005). An LRRK2 mutation as a cause for the parkinsonism in the original PARK8 family. *Annals of Neurology*, 57(6), 918-921. doi:10.1002/ ana.20484
- Games, D., Adams, D., Alessandrini, R., Barbour, R., Berthelette, P., Blackwell, C.,

. . . . et al. (1995). Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. *Nature*, *373*(6514), 523-527. doi:10.1038/373523a0

- Gessel, M. M., Bernstein, S., Kemper, M., Teplow, D. B., & Bowers, M. T. (2012). Familial Alzheimer's disease mutations differentially alter amyloid β-protein oligomerization. ACS Chemical Neuroscience, 3(11), 909-918. doi:10.1021/cn300050d
- Gilks, W. P., Abou-Sleiman, P. M., Gandhi, S., Jain, S., Singleton, A., Lees, A. J., . . . Wood, N. W. (2005). Common LRRK2 mutation in idiopathic Parkinson's disease. *Lancet*, 365(9457), 415-416. doi:10.1016/ s0140-6736(05)17830-1
- Hamm, V., Heraud, C., Bott, J.-B., Herbeaux, K., Strittmatter, C., Mathis, C., & Goutagny, R. (2017). Differential contribution of APP metabolites to early cognitive deficits in a TgCRND8 mouse model of Alzheimer's disease. *Science Advances*, 3(2). doi:10.1126/sciadv.1601068
- Hattori, N., Matsumine, H., Asakawa, S., Kitada, T., Yoshino, H., Elibol, B., . . . Mizuno, Y. (1998). Point mutations (Thr240Arg and Ala311Stop) in the Parkin gene. *Biochemical and Biophysical Research Communications, 249*(3), 754-758. doi:10.1006/bbrc.1998.9134
- Healy, D. G., Abou-Sleiman, P. M., Ahmadi, K. R., Muqit, M. M. K., Bhatia, K. P., Quinn, N. P., . . . Wood, N. W. (2004). The gene responsible for PARK6 Parkinson's disease, PINK1, does not influence common forms of parkinsonism. *Annals of Neurology*, 56(3), 329-335. doi:10.1002/ana.20206
- Hinkle, K. M., Yue, M., Behrouz, B., Dächsel, J. C., Lincoln, S. J., Bowles, E. E., . . . Melrose, H. L. (2012). LRRK2 knockout mice have an intact dopaminergic system but display alterations in exploratory and motor co-ordination behaviors. *Molecular Neurodegeneration*, 7, 25. doi:10.1186/1750-1326-7-25
- Hollingworth, P., Harold, D., Sims, R., Gerrish, A., Lambert, J. C., Carrasquillo, M. M., . . . Williams, J. (2011). Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with

Alzheimer's disease. *Nature genetics,* 43(5), 429-435. doi:10.1038/ng.803

- Hor, J. H., Soh, E. S. Y., Tan, L. Y., Lim, V. J. W., Santosa, M. M., Winanto, . . . Ng, S. Y. (2018). Cell cycle inhibitors protect motor neurons in an organoid model of Spinal Muscular Atrophy. *Cell Death & Disease*, 9. doi:10.1038/s41419-018-1081-0
- Israel, M. A., Yuan, S. H., Bardy, C., Reyna, S. M., Mu, Y., Herrera, C., . . . Goldstein, L. S. B. (2012). Probing sporadic and familial Alzheimer's disease using induced pluripotent stem cells. *Nature*, 482(7384), 216-U107. doi:10.1038/nature10821
- Iwatsubo, T., Odaka, A., Suzuki, N., Mizusawa, H., Nukina, N., & Ihara, Y. (1994). VISUALIZATION OF A-BETA-42(43) AND A-BETA-40 IN SENILE PLAQUES WITH END-SPECIFIC A-BETA MONOCLONALS - EVIDENCE THAT AN INITIALLY DEPOSITED SPECIES IS A-BETA-42(43). *Neuron*, *13*(1), 45-53. doi:10.1016/0896-6273(94)90458-8
- Jo, J., Xiao, Y. X., Sun, A. X., Cukuroglu, E., Tran, H. D., Goke, J., ... Ng, H. H. (2016). Midbrain-like Organoids from Human Pluripotent Stem Cells Contain Functional Dopaminergic and Neuromelanin-Producing Neurons. Cell Stem Cell, 19(2), 248-257. doi:10.1016/j.stem.2016.07.005
- Kawada, J., Kaneda, S., Kirihara, T., Maroof, A., Levi, T., Eggan, K., . . . Ikeuchi, Y. (2017). Generation of a Motor Nerve Organoid with Human Stem Cell-Derived Neurons. *Stem Cell Reports*, 9(5), 1441-1449. doi:10.1016/j.stemcr.2017.09.021
- Kitada, T., Asakawa, S., Hattori, N., Matsumine, H., Yamamura, Y., Minoshima, S., . . . Shimizu, N. (1998). Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature*, 392(6676), 605-608. doi:10.1038/33416
- Klein, C., Djarmati, A., Hedrich, K., Schafer, N., Scaglione, C., Marchese, R., . . . Pramstaller, P. P. (2005). PINK1, Parkin, and DJ-1 mutations in Italian patients with early-onset parkinsonism. *European Journal of Human Genetics*, *13*(9), 1086-1093. doi:10.1038/sj.ejhg.5201455
- Klein, H. U., McCabe, C., Gjoneska, E., Sullivan, S. E., Kaskow, B. J., Tang, A., . . . De Jager, P. L. (2019). Epigenome-wide study uncovers large-scale changes in histone

acetylation driven by tau pathology in aging and Alzheimer's human brains. *Nature Neuroscience*, 22(1), 37-46. doi:10.1038/s41593-018-0291-1

- Lancaster, M. A., Renner, M., Martin, C.-A., Wenzel, D., Bicknell, L. S., Hurles, M. E., . . . Knoblich, J. A. (2013). Cerebral organoids model human brain development and microcephaly. *Nature*, 501(7467), 373-+. doi:10.1038/nature12517
- Lee, G., Papapetrou, E. P., Kim, H., Chambers, S. M., Tomishima, M. J., Fasano, C. A., ... Studer, L. (2009). Modelling pathogenesis and treatment of familial dysautonomia using patient-specific iPSCs. *Nature*, 461(7262), 402-U100. doi:10.1038/nature08320
- Lewis, J., Dickson, D. W., Lin, W. L., Chisholm, L., Corral, A., Jones, G., . . . McGowan, E. (2001). Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science*, 293(5534), 1487-1491. doi:10.1126/science.1058189
- Lewis, J., McGowan, E., Rockwood, J., Melrose, H., Nacharaju, P., Van Slegtenhorst, M., . . . Hutton, M. (2000). Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nature genetics*, 25(4), 402-405. doi:10.1038/78078
- Lopes, K. P., Snijders, G. J. L., Humphrey, J., Allan, A., Sneeboer, M. A. M., Navarro, E., . . . Raj, T. (2022). Genetic analysis of the human microglial transcriptome across brain regions, aging and disease pathologies. *Nature genetics*, 54(1), 4-17. doi:10.1038/s41588-021-00976-y
- Lucking, C. B., Durr, A., Bonifati, V., Vaughan, J., De Michele, G., Gasser, T., . . . French Parkinsons Dis Genetics, S. (2000). Association between early-onset Parkinson's disease and mutations in the parkin gene. *New England Journal of Medicine*, *342*(21), 1560-1567. doi:10.1056/nejm200005253422103
- Ma, C., Bao, A. M., & Yan, X. X. (2019). Significance of human brain banking and recent advances in China and foreign countries. *Journal of Chongqing Medical University*, 44(04), 547-550. doi:10.13406/j.cnki.cyxb.002068
- Ma, C., Bao, A. M., Yan, X. X., & Swaab, D. F. (2019). Progress in Human Brain Banking in China. *Neuroscience Bulletin*, 35(2), 179-182. doi:10.1007/s12264-019-00350-3

- Ma, Y., Dammer, E. B., Felsky, D., Duong, D. M., Klein, H. U., White, C. C., . . . De Jager, P. L. (2021). Atlas of RNA editing events affecting protein expression in aged and Alzheimer's disease human brain tissue. *Nature Communications, 12*(1), 7035. doi:10.1038/s41467-021-27204-9
- Macpherson, T. A., Garver, K. L., Turner, J. H., Diggans, G. R., Marchese, S. G., & Poole, G. C. (1985). Predicting in vitro tissue culture growth for cytogenetic evaluation of stillborn fetuses. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 19(3), 167-174. doi:10.1016/0028-2243(85)90151-0
- Marchetto, M. C. N., Carromeu, C., Acab, A., Yu, D., Yeo, G. W., Mu, Y., . . . Muotri, A. R. (2010). A Model for Neural Development and Treatment of Rett Syndrome Using Human Induced Pluripotent Stem Cells. *Cell*, 143(4), 527-539. <u>doi:10.1016/j.cell.2010.10.016</u>
- Mariani, J., Coppola, G., Zhang, P., Abyzov, A., Provini, L., Tomasini, L., . . . Vaccarino, F. M. (2015). FOXG1-Dependent Dysregulation of GABA/Glutamate Neuron Differentiation in Autism Spectrum Disorders. *Cell*, 162(2), 375-390. doi:10.1016/j.cell.2015.06.034
- Maruyama, M., Shimada, H., Suhara, T., Shinotoh, H., Ji, B., Maeda, J., . . . Higuchi, M. (2013). Imaging of tau pathology in a tauopathy mouse model and in Alzheimer patients compared to normal controls. *Neuron*, 79(6), 1094-1108. <u>doi:10.1016/j.</u> <u>neuron.2013.07.037</u>
- McInerney-Leo, A., Hadley, D. W., Gwinn-Hardy, K., & Hardy, J. (2005). Genetic testing in Parkinson's disease. *Movement Disorders*, 20(1), 1-10. doi:10.1002/mds. 20316
- Mellios, N., Feldman, D. A., Sheridan, S. D., Ip, J. P. K., Kwok, S., Amoah, S. K., ... Sur, M. (2018). MeCP2-regulated miRNAs control early human neurogenesis through differential effects on ERK and AKT signaling. *Molecular Psychiatry*, 23(4), 1051-1065. doi:10.1038/mp.2017.86
- Metzger, J. M., & Emborg, M. E. (2019). Autonomic dysfunction in Parkinson disease and animal models. *Clinical Autonomic Research*, 29(4), 397-414. doi:10.1007/ s10286-018-00584-7

- Miller, J. D., Ganat, Y. M., Kishinevsky, S., Bowman, R. L., Liu, B., Tu, E. Y., . . . Studer, L. (2013). Human iPSC-Based Modeling of Late-Onset Disease via Progerin-Induced Aging. *Cell Stem Cell, 13*(6), 691-705. doi:10.1016/j. stem.2013.11.006
- Montine, T. J., Phelps, C. H., Beach, T. G., Bigio,
 E. H., Cairns, N. J., Dickson, D. W., . . .
 Hyman, B. T. (2012). National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathologica*, 123(1), 1-11. doi:10.1007/s00401-011-0910-3
- Monzel, A. S., Smits, L. M., Hemmer, K., Hachi, S., Moreno, E. L., van Wuellen, T., . . . Schwamborn, J. C. (2017). Derivation of Human Midbrain-Specific Organoids from Neuroepithelial Stem Cells. *Stem Cell Reports*, 8(5), 1144-1154. <u>doi:10.1016/j.</u> <u>stemcr.2017.03.010</u>
- Moore, K. A., Kohno, T., Karchewski, L. A., Scholz, J., Baba, H., & Woolf, C. J. (2002). Partial peripheral nerve injury promotes a selective loss of GABAergic inhibition in the superficial dorsal horn of the spinal cord. *Journal of Neuroscience*, 22(15), 6724-6731. <u>doi:10.1523/</u> jneurosci.22-15-06724.2002
- Morris, R. G., & Salmon, D. P. (2007). The centennial of Alzheimer's disease and the publication of "Uber eine eigenartige Erkankung der Hirnrinde" by Alöis Alzheimer. *Cortex*, 43(7), 821-825. doi:10.1016/s0010-9452(08)70681-6
- Muguruma, K., Nishiyama, A., Ono, Y., Miyawaki, H., Mizuhara, E., Hori, S., . . . Sasai, Y. (2010). Ontogeny-recapitulating generation and tissue integration of ES cell-derived Purkinje cells. *Nature Neuroscience*, *13*(10), 1171-1180. doi:10.1038/nn.2638
- Munoz, E., Oliva, R., Obach, V., Marti, M. J., Pastor, P., Ballesta, F., & Tolosa, E. (1997). Identification of Spanish familial Parkinson's disease and screening for the Ala53Thr mutation of the alpha-synuclein gene in early onset patients. *Neuroscience Letters*, 235(1-2), 57-60. doi:10.1016/s0304-3940(97)00710-6
- Naj, A. C., Jun, G., Beecham, G. W., Wang, L.-S., Vardarajan, B. N., Buros, J., . . .

Schellenberg, G. D. (2011). Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nature genetics*, *43*(5), 436-441. doi:10.1038/ ng.801

- Nakano, T., Ando, S., Takata, N., Kawada, M., Muguruma, K., Sekiguchi, K., . . . Sasai, Y. (2012). Self-Formation of Optic Cups and Storable Stratified Neural Retina from Human ESCs. *Cell Stem Cell*, 10(6), 771-785. doi:10.1016/j.stem.2012.05.009
- Nativio, R., Lan, Y., Donahue, G., Sidoli, S., Berson, A., Srinivasan, A. R., . . . Berger, S. L. (2020). An integrated multi-omics approach identifies epigenetic alterations associated with Alzheimer's disease. *Nature genetics*, 52(10), 1024-1035. doi:10.1038/s41588-020-0696-0
- Nichols, W. C., Pankratz, N., Hernandez, D., Paisan-Ruiz, C., Jain, S., Halter, C. A., ... Parkinson Study Grp, P. I. (2005). Genetic screening for a single common LRRK2 mutation in familial Parkinson's disease. *Lancet*, 365(9457), 410-412. doi:10.1016/ s0140-6736(05)17828-3
- Nussbaum, R. L., & Polymeropoulos, M. H. (1997). Genetics of Parkinson's disease. *Human Molecular Genetics*, 6(10), 1687-1691. doi:10.1093/hmg/6.10.1687
- Pang, K. L., Jiang, R. C., Zhang, W., Yang, Z. Y., Li, L. L., Shimozawa, M., . . . Lu, B. (2022). An App knock-in rat model for Alzheimer's disease exhibiting A beta and tau pathologies, neuronal death and cognitive impairments. *Cell Research*, 32(2), 157-175. doi:10.1038/s41422-021-00582-x
- Panitch, R., Hu, J., Chung, J., Zhu, C., Meng, G., Xia, W., . . . Jun, G. R. (2021). Integrative brain transcriptome analysis links complement component 4 and HSPA2 to the APOE ε2 protective effect in Alzheimer disease. *Molecular Psychiatry*, 26(10), 6054-6064. doi:10.1038/s41380-021-01266-z
- Pasca, A. M., Sloan, S. A., Clarke, L. E., Tian, Y., Makinson, C. D., Huber, N., . . . Pasca, S. P. (2015). Functional cortical neurons and astrocytes from human pluripotent stem cells in 3D culture. *Nature Methods*, *12*(7), 671-+. doi:10.1038/nmeth.3415
- Pringsheim, T., Jette, N., Frolkis, A., & Steeves, T. D. (2014). The prevalence of Parkinson's

disease: a systematic review and metaanalysis. *Movement Disorders*, 29(13), 1583-1590. doi:10.1002/mds.25945

- Qi, Y. J., Lu, Y. R., Shi, L. G., Demmers, J. A. A., Bezstarosti, K., Rijkers, E., . . . Bao, A. M. (2022). Distinct proteomic profiles in prefrontal subareas of elderly major depressive disorder and bipolar disorder patients. *Translational Psychiatry*, 12(1). doi:10.1038/s41398-022-02040-7
- Qian, X., Ha Nam, N., Song, M. M., Hadiono, C., Ogden, S. C., Hammack, C., ... Ming, G.-I. (2016). Brain-Region-Specific Organoids Using Mini-bioreactors for Modeling ZIKV Exposure. *Cell*, 165(5), 1238-1254. doi:10.1016/j.cell.2016.04.032
- Qian, X. Y., Jacob, F., Song, M. M., Nguyen, H. N., Song, H. J., & Ming, G. L. (2018). Generation of human brain region-specific organoids using a miniaturized spinning bioreactor. *Nature Protocols*, *13*(3), 565-580. doi:10.1038/nprot.2017.152
- Qiu, W., Zhang, H., Bao, A., Zhu, K., Huang, Y., Yan, X., . . . Ma, C. (2019). Standardized Operational Protocol for Human Brain Banking in China. *Neuroscience Bulletin*, *35*(2), 270-276. <u>doi:10.1007/s12264-018-0306-7</u>
- Rademaker, M. C., de Lange, G. M., & Palmen, S. J. M. C. (2018). Chapter 1 - The Netherlands Brain Bank for Psychiatry. In I. Huitinga & M. J. Webster (Eds.), *Handbook of Clinical Neurology* (Vol. 150, pp. 3-16): Elsevier.
- Rathke-Hartlieb, S., Kahle, P. J., Neumann, M., Ozmen, L., Haid, S., Okochi, M., . . . Schulz, J. B. (2001). Sensitivity to MPTP is not increased in Parkinson's disease-associated mutant alphasynuclein transgenic mice. *Journal* of *Neurochemistry*, 77(4), 1181-1184. doi:10.1046/j.1471-4159.2001.00366.x
- Rodriguez-Oroz, M. C., Jahanshahi, M., Krack, P., Litvan, I., Macias, R., Bezard, E., & Obeso, J. A. (2009). Initial clinical manifestations of Parkinson's disease: features and pathophysiological mechanisms. *Lancet Neurology*, 8(12), 1128-1139. doi:10.1016/ s1474-4422(09)70293-5
- Rogaeva, E., Johnson, J., Lang, A. E., Gulick, C., Gwinn-Hardy, K., Kawarai, T., . . . Singleton, A. B. (2004). Analysis of the PINK1 gene in a large cohort of cases with

- Sakaguchi, H., Kadoshima, T., Soen, M., Narii, N., Ishida, Y., Ohgushi, M., . . . Sasai, Y. (2015). Generation of functional hippocampal neurons from self-organizing human embryonic stem cell-derived dorsomedial telencephalic tissue. *Nature Communications*, <u>6. doi:10.1038/ncomms9896</u>
- Samarasekera, N., Al-Shahi Salman, R., Huitinga, I., Klioueva, N., McLean, C. A., Kretzschmar, H., ... Ironside, J. W. (2013). Brain banking for neurological disorders. *Lancet Neurology*, 12(11), 1096-1105. doi:10.1016/s1474-4422(13)70202-3
- Samarasekera, N., Salman, R. A.-S., Huitinga, I., Klioueva, N., McLean, C. A., Kretzschmar, H., . . Ironside, J. W. (2013). Brain banking for neurological disorders. *Lancet Neurology*, *12*(11), 1096-1105. doi:10.1016/s1474-4422(13)70202-3
- Sanchez-Varo, R., Mejias-Ortega, M., Fernandez-Valenzuela, J. J., Nuñez-Diaz, C., Caceres-Palomo, L., Vegas-Gomez, L., . . . Gutierrez, A. (2022). Transgenic Mouse Models of Alzheimer's Disease: An Integrative Analysis. *international journal of molecular sciences*, 23(10). doi:10.3390/ijms23105404
- Sanchez, G., Varaschin, R. K., Büeler, H., Marcogliese, P. C., Park, D. S., & Trudeau, L. E. (2014). Unaltered striatal dopamine release levels in young Parkin knockout, Pink1 knockout, DJ-1 knockout and LRRK2 R1441G transgenic mice. *PLoS One*, 9(4), e94826. doi:10.1371/journal. pone.0094826
- Sasaguri, H., Nilsson, P., Hashimoto, S., Nagata, K., Saito, T., De Strooper, B., . . . Saido, T. C. (2017). APP mouse models for Alzheimer's disease preclinical studies. *Embo Journal*, 36(17), 2473-2487. doi:10.15252/embj.201797397
- Soldner, F., Hockemeyer, D., Beard, C., Gao, Q., Bell, G. W., Cook, E. G., . . . Jaenisch, R. (2009). Parkinson's Disease Patient-Derived Induced Pluripotent Stem Cells Free of Viral Reprogramming Factors. *Cell*, *136*(5), 964-977. doi:10.1016/j.cell.2009.02.013
- StGeorge-Hyslop, P. H., Tanzi, R. E., Polinsky, R. J., Haines, J. L., Nee, L., Watkins, P. C., . . . et al. (1987). The genetic defect causing

familial Alzheimer's disease maps on chromosome 21. *Science*, 235(4791), 885-890. doi:10.1126/science.2880399

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- Stachowiak, E. K., Benson, C. A., Narla, S. T., Dimitri, A., Chuye, L. E. B., Dhiman, S.,
 Stachowiak, M. K. (2017).
 Cerebral organoids reveal early cortical maldevelopment in schizophreniacomputational anatomy and genomics, role of FGFR1. *Translational Psychiatry*, 7. doi:10.1038/s41398-017-0054-x
- Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K., & Yamanaka, S. (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*, 131(5), 861-872. doi:10.1016/j.cell.2007.11.019
- Toker, L., Mancarci, B. O., Tripathy, S., & Pavlidis, P. (2018). Transcriptomic Evidence for Alterations in Astrocytes and Parvalbumin Interneurons in Subjects With Bipolar Disorder and Schizophrenia. *Bbiological Psychiatry*, 84(11), 787-796. doi:10.1016/j.biopsych.2018.07.010
- Valente, E. M., Abou-Sleiman, P. M., Caputo, V., Muqit, M. M. K., Harvey, K., Gispert, S., . . . Wood, N. W. (2004). Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science*, 304(5674), 1158-1160. doi:10.1126/science.1096284
- Vallortigara, J., Rangarajan, S., Whitfield, D., Alghamdi, A., Howlett, D., Hortobágyi, T.,
 . Francis, P. (2014). Dynamin1 concentration in the prefrontal cortex is associated with cognitive impairment in Lewy body dementia. *F1000Research*, *3*, 108. doi:10.12688/f1000research.3786.1
- Vallortigara, J., Whitfield, D., Quelch, W., Alghamdi, A., Howlett, D., Hortobágyi, T., . . . Francis, P. T. (2016). Decreased Levels of VAMP2 and Monomeric Alpha-Synuclein Correlate with Duration of Dementia. *Journal of Alzheimers Disease*, 50(1), 101-110. doi:10.3233/jad-150707
- Vialle, R. A., de Paiva Lopes, K., Bennett, D. A., Crary, J. F., & Raj, T. (2022). Integrating whole-genome sequencing with multiomic data reveals the impact of structural variants on gene regulation in the human brain. *Nature Neuroscience*, *25*(4), 504-514. doi:10.1038/s41593-022-01031-7
- Wang, L., Xia, Y., Chen, Y., Dai, R., Qiu, W., Meng, Q.,... Chen, C. (2019). Brain Banks

Spur New Frontiers in Neuropsychiatric Research and Strategies for Analysis and Validation. *Genomics Proteomics* & *Bioinformatics*, 17(4), 402-414. doi:10.1016/j.gpb.2019.02.002

- Wang, Q., Zhang, Y., Wang, M., Song, W.M., Shen, Q., McKenzie, A., . . . Zhang, B. (2019). The landscape of multiscale transcriptomic networks and key regulators in Parkinson's disease. *Nature Communications*, 10(1), 5234. doi:10.1038/s41467-019-13144-y
- Webster, S. J., Bachstetter, A. D., Nelson, P. T., Schmitt, F. A., & Van Eldik, L. J. (2014). Using mice to modelAlzheimer's dementia: an overview of the clinical disease and the preclinical behavioral changes in 10 mouse models. *Frontiers in Genetics*, 5, 88. doi:10.3389/fgene.2014.00088
- Whitfield, D. R., Vallortigara, J., Alghamdi, A., Howlett, D., Hortobágyi, T., Johnson, M., . . . Francis, P. T. (2014). Assessment of ZnT3 and PSD95 protein levels in Lewy body dementias and Alzheimer's disease: association with cognitive impairment. *Neurobiology of Aging*, 35(12), 2836-2844. doi:10.1016/j.neurobiolaging.2014.06.015
- Xiong, F., Ge, W., & Ma, C. (2019). Quantitative proteomics reveals distinct composition of

amyloid plaques in Alzheimer's disease. *Alzheimers & Dementia*, 15(3), 429-440. doi:10.1016/j.jalz.2018.10.006

- Zareparsi, S., Kay, J., Camicioli, R., Kramer, P., Nutt, J., Bird, T., . . . Payami, H. (1998). Analysis of the alpha-synuclein G209A mutation in familial Parkinson's disease. *Lancet*, 351(9095), 37-38. doi:10.1016/ s0140-6736(05)78089-2
- Zhang, Y., Pak, C., Han, Y., Ahlenius, H., Zhang,
 Z., Chanda, S., . . . Suedhof, T. C. (2013).
 Rapid Single-Step Induction of Functional Neurons from Human Pluripotent Stem Cells. *Neuron*, 78(5), 785-798. doi:10.1016/j.neuron.2013.05.029
- Zimprich, A., Biskup, S., Leitner, P., Lichtner, P., Farrer, M., Lincoln, S., ... Gasser, T. (2004). Mutations in LRRK2 cause autosomaldominant Parkinsonism with pleomorphic pathology. *Neuron*, 44(4), 601-607. doi:10.1016/j.neuron.2004.11.005
- Zou, F., Chai, H. S., Younkin, C. S., Allen, M., Crook, J., Pankratz, V. S., ... Ertekin-Taner, N. (2012). Brain expression genome-wide association study (eGWAS) identifies human disease-associated variants. *PLoS Genetics*, 8(6), e1002707. doi:10.1371/ journal.pgen.1002707



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