

Roles of microglial calcium channels in neurodegenerative diseases

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Abstract: Microglia are crucial for neurodevelopment, and the maintenance of central nervous system functions. Calcium signals in microglia regulate the neuronal plasticity critical for learning, memory, and neuron survival. Growing evidence highlights the pivotal function of calcium channels in microglia, along with their cognate proteins, in modulating oxidative stress, neuroinflammation, and multiple neurodegenerative diseases such as Alzheimer's disease and Parkinson's disease. In this review, we summarized the important role and regulatory mechanism of these critical calcium channels and their associated proteins, highlighting their potential as novel therapeutic targets for neurodegenerative diseases.

Keywords: Calcium channels; Microglia; Nervous system; Neurological disorders.

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INTRODUCTION

Microglial cells, serving as the intrinsic immune constituents of the central nervous system (CNS), originate from erythromyeloid progenitor cells in the embryonic yolk sac. They migrate to the CNS during embryogenesis, afterwards, they proliferate and extend throughout the brain parenchyma¹. Within the cerebral framework, microglia represent the predominant immune cellular subtype, constituting more than 80% of the entire immune cell population². Microglia play a crucial role in cerebral development, monitoring neuronal activity, and modulating learning and memory processes. Meanwhile, they also function as intrinsic phagocytes and sentinels of injury within the brain parenchyma^{3,4}. Immune modulators produced by microglia extend to damaged areas and phagocytize cell debris, apoptotic neurons, and synapses. Specifically, microglial activation instigates the secretion of cytokines and chemokines. This process is concomitant with an elevation in intracellular calcium concentrations⁵. Microglia are activated in response to pathogenic insults, marked by alternated gene expression and cellular function. Advanced investigations employing single-cell RNA sequencing elucidate the continuous change of microglia correlating with disease progression. Microglia is highly heterogeneous. A recent study identifies a disease-associated microglial (DAM) signature, hallmarked with the increase of a spectrum of genes, such as Triggering Receptor Expressed on Myeloid Cells 2 (Trem2) and Apolipoprotein E (ApoE)^{6,7}.

Upon activation, microglia exhibit two distinct polarization patterns: pro-inflammatory and anti-inflammatory. Pro-inflammatory microglia release pro-inflammatory cytokines and chemokines,

whereas anti-inflammatory microglia enhance phagocytosis and secrete cytokines that facilitate tissue repair and neuroprotection⁸. Moreover, imbalanced microglial function has been suggested as a major contributor to various neurological diseases⁹⁻¹¹. Activation of microglial cells facilitates calcium permeability of the cellular membrane, thereby enhancing the calcium influx into the cytoplasm^{12,13}. This transient overload of intracellular calcium concentration is imperative for the activation of microglia¹⁴.

Modifications in the ionic homeostasis of microglia about sodium (Na⁺), calcium (Ca²⁺), potassium (K⁺), hydrogen (H⁺), and chloride (Cl⁻) ions are essential for the activation of microglia. This activation encompasses several physiological processes such as the release of cytokines, cellular migration, proliferation, and the synthesis of reactive oxygen species (ROS). Calcium signaling is particularly salient in the initiation of microglial activation and its related physiological responses¹⁵. Microgliosis and pathological remodeling of these cells are largely under the control of calcium signaling in microglia¹⁶. G protein-coupled receptors, ion channels, calcium-binding proteins, and ion exchangers are key regulators of calcium transportation through the membrane of cytoplasm, mitochondria, and the ER¹³.

Activated microglia can be observed in chronic degenerative diseases, including Alzheimer's disease (AD) and Parkinson's disease (PD). Neurodegenerative diseases are marked by a spectrum of cognitive and behavioral deficits that are attributable to insidious and progressive degeneration of neurons¹⁷. Aging significantly predisposes individuals to the development and exacerbation of these disorders, with data indicating that upwards of 90% of neurodegenerative cases are diagnosed in persons beyond the age of 60¹⁸. Additionally, aging causes microglia to respond to external stimuli more strongly¹⁹. The dysregulation of microglial transitional responses is thought to be the cause of persistent inflammation in the aging cerebral environment²⁰.

Intracellular calcium dysregulation is commonly observed during aging and neurodegeneration. Disruption in microglial calcium signaling has been implicated in the pathogenesis of several CNS disorders²¹. Cellular processes that contribute to the homeostasis of calcium are also disrupted in neurodegenerative diseases. Oxidative stress, disordered energy metabolism, and

modifications in disease-related proteins all contribute to calcium-dependent synaptic dysfunction. Altered calcium homeostasis plays a critical role in the long-term neurodegenerative processes that contribute to both AD, PD, and other inflammation-mediated neural damage²². The role and regulatory role of microglial intracellular ionic homeostasis in neurodegenerative diseases are still unclear.

This review synthesizes recent developments in how calcium homeostasis contributing to the development neurodegenerative diseases while highlighting the potential emergent therapeutic strategies aiming at calcium homeostasis in the treatment of those diseases.

Ca²⁺ HOMEOSTASIS IN MICROGLIAL FUNCTION

Ca²⁺, stored in the ER and mitochondria, serves as a vital secondary messenger, functioning both intracellularly and intercellularly²³. Ca²⁺ functions as information-carrying messengers due to their reversible complexation with proteins²⁴. Intracellular Ca²⁺ is implicated in various signaling activities such as enzyme activation, exocytosis, gene expression, programmed cell death, synaptic communication, and regulation of ROS²⁵. Intracellular calcium signaling in neurons is pivotal for neural differentiation, the consolidation of memory, and the regulation of synaptic plasticity. Similarly, dysregulation of calcium homeostasis can induce neuronal damage and cell death²². Under resting state, the intracellular Ca²⁺ level is maintained at a low level (~10⁻⁷ M)²⁶. Upon activation, Ca²⁺ influx is tightly controlled by complex molecular regulatory mechanisms.

Many microglial functions are mediated by calcium ions²⁷⁻²⁹. Specifically, the activation of microglia is associated with an elevation of intracellular calcium levels, a cascade that is essential for triggering the secretion of cytokines and chemokines⁵. Elevated intracellular calcium levels initiate signaling cascades, which are crucial for biological processes, including the activation of enzymatic pathways regulating gene transcription, cell proliferation, differentiation, and migration³⁰. Normally, microglia are considered non-excitable cells¹³. Microglial intracellular calcium signaling is modulated by electrochemical calcium influx through channels and receptors, and efflux via various calcium pumps and exchangers against the concentration

gradient³¹. The differential distribution of these channels and transporters allows distinct intracellular compartments to manage calcium uniquely. In microglia, calcium channels and transporters facilitate the entry and exit of Ca²⁺, preserving intracellular Ca²⁺ balance¹³.

Recent studies have revealed that dysregulated calcium signaling is associated with a spectrum of neurodegenerative diseases, such as AD, PD, Amyotrophic lateral sclerosis (ALS), and Huntington's disease (HD)^{32,33}. This is connected with changes in calcium ion buffering capacity, activity of calcium channels and other calcium regulator proteins^{34,35}, and impaired mitochondrial and ER calcium

processing³⁶, disrupted energy metabolism, and oxidative stress²².

TYPES OF Ca²⁺ CHANNELS IN MICROGLIA

On the cytoplasmatic membrane of microglial cells, there are multiple types of calcium channels and calcium-related proteins, including store-operated Ca²⁺ entry (SOCE) channels, transient receptor potential (TRP) channels, voltage-gated Ca²⁺ channels (VGCCs), calcium homeostasis modulator family protein 2 (Calhm2), and Na⁺ /Ca²⁺ exchanger (NCX) (Figure 1).

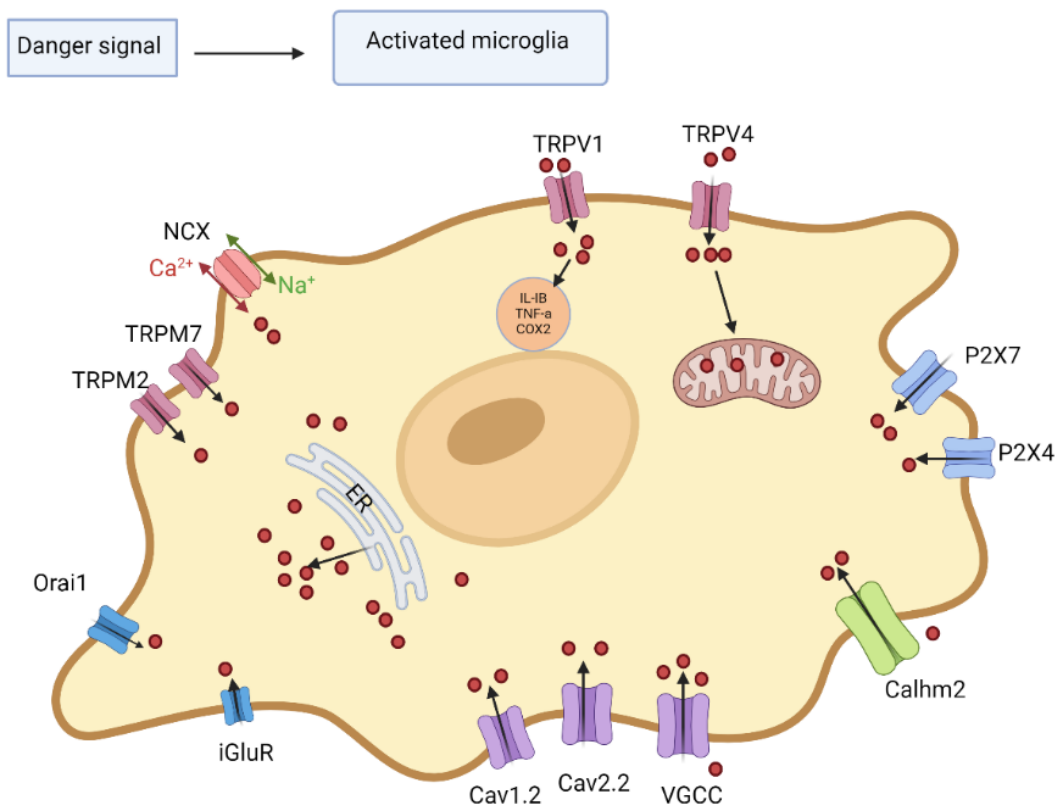


Figure 1. Activated microglial cell calcium channels and their related proteins regulating ion homeostasis mechanism in pathologic conditions. Schematic drawing of a microglial cell, illustrating receptors/channels known to increase calcium channels during pathologic conditions. Ca²⁺ entry pathway: Store-operated Ca²⁺ entry channels, Orai1; Transient receptor potential channel, TRPM2, TRPM7, TPPV1, TRPV4; Voltage-gated Ca²⁺ channels, Cav1.2, Cav2.2; The calcium homeostatic regulatory protein, calhm2. Receptor-operated Ca²⁺ channels, P2X4, P2X7; Bidirectional ion transporter: Sodium-calcium exchanger (NCX), facilitating the exchange of Ca²⁺ efflux for Na⁺ influx.

SOCE channels: Microglial cells exhibit high levels of SOCE, mediated by stromal interaction molecules (STIM1/2) and Orai plasma-membrane pore-forming subunits (Orai1/2/3)³⁷. Purines are

released from damaged brain cells, which stimulate microglia to migrate to injury sites and initiate phagocytosis. These processes are regulated by calcium-dependent purinergic signaling pathways,

involving either P2Y receptor-induced internal calcium release or P2X receptor-mediated plasmalemmal calcium influx³⁸. Interestingly, it is reported that SOCE contributes to the purinergic activation in microglia^{37,39}.

TRP channel: TRP channels are typical non-selective cation channels, participating in calcium transportation across the plasma membrane and ER, thus maintaining microglial calcium homeostasis and dynamics^{40,41}. TRP channels are ubiquitously present within the CNS, contributing to the regulation of intracellular calcium concentration ($[Ca^{2+}]_i$) by facilitating calcium influx through the cytoplasmic membrane and calcium release from intracellular stores⁴². Based on sequence homology, the 28 mammalian cation channels in the TRP family are categorized into six subfamilies: TRPC, TRPM, TRPV, TRPA, TRPP, and TRPML^{43,44}. Specifically, TRPV, TRPM, and TRPC channels, located in microglia, participate in diverse functions such as osmotic regulation, cytokine production, cell proliferation, death, microgliosis, and oxidative stress response⁴³. TRPV1 activity in microglia is primarily linked to neurotoxicity via pro-inflammatory cytokine production and induction of oxidative stress⁴⁴. TRPV1, mainly present in intracellular organelles such as mitochondria, significantly contributes to triggering microglial migration⁴⁵. TRPV1 activation elicits an elevation in mitochondrial calcium levels and induces membrane depolarization, subsequently leading to an upsurge in ROS generation⁴⁶.

VGCCs: VGCCs are commonly expressed on excitable cells. It contains several subunits, the $\alpha 1$, $\alpha 2/\delta$, and β subunits^{47,48}. Among these subunits, the $\alpha 1$ subunit is the principal component, containing the voltage-sensing domains and defining the unique properties characteristic of each VGCC subtype^{47,49-52}. Voltage-gated calcium channels (VGCCs) are categorized into various families, such as Cav1, Cav2, and Cav3, each comprising multiple subtypes⁵³. In diverse cellular contexts, the entry of Ca^{2+} into the cytosol through VGCCs regulates the enzymatic activity, gene expression, and a variety of biochemical mechanisms. The Cav1 calcium channel family is comprised of four distinct subfamilies, named Cav1.1 to Cav1.4, each of which exhibits a characterized sensitivity to calcium channel antagonists⁵⁴. Among the Cav1 family, Cav1.3 has been reported to be associated with the pathophysiological mechanisms underlying PD. The Cav1.3 channel is reported to play a critical

role in stabilizing pacemaker currents in substantia nigra pars compacta (SNc) dopaminergic neurons⁵⁵, and pharmacological blockade of this channel inhibits neuronal apoptosis in the SNc, and improves behavioral deficiencies in PD murine models⁵⁶. Additionally, microglia have been reported to express Cav1.2 and Cav2.2 channel subtypes⁵³. VGCCs remain closed at physiological or resting membrane potential and activate upon membrane depolarization⁵⁷. The Cav family translates alterations in the cell surface membrane potential into localized increases in $[Ca^{2+}]_i$ ⁵⁸.

Calhm2: The calcium homeostatic regulatory protein family (CALHMs) constitutes voltage-gated non-selective ion channels that play a crucial role in taste signaling and attenuating neuronal toxicity by facilitating major adenosine triphosphate (ATP) release⁵⁹. The CALHMs contain 6 identified members (Calhm1 to Calhm6)⁵⁹. The association between CALHMs and neurodegenerative disease research has recently drawn wide attention. Calhm1 encodes a 346-amino acid protein that regulates the permeability of calcium ions across the plasma membrane. The Calhm1 gene harbors a nonsynonymous polymorphism, P86L, which has been linked to AD development, potentially through the alteration of β -amyloid levels⁶⁰. Moreover, Calhm1 is reported to express on type II taste bud cells and contributes to the release of ATP through a voltage-gated channel and affects taste perception⁶¹. Calhm1 regulates calcium homeostasis, influences the production of amyloid beta ($A\beta$), and modulates neuronal cell susceptibility to toxicity induced by $A\beta$ ⁶². Calhm1 expression is observed in murine cerebral tissues, yet it is absent in human cerebral samples. Moreover, calhm1 knockout mice do not exhibit any discernible cognitive abnormalities⁶³. Calhm3 has been shown to form an isomeric channel with Calhm1, and taste-evoked ATP release was eliminated when Calhm3 was deleted⁶⁴. Under cryo-EM conditions, Calhm2 is observed to be a quadruple transmembrane protein, and Calhm2 channels can form both gap junctions and undecameric channels⁶⁵. It has been discovered that Calhm2 mediates the transfer of calcium and ATP, controlling calcium and ATP concentrations within and outside of cells^{61,63-67}. It is important to note that the functional and biological significance of the remaining CALHM family members, specifically Calhm4 to Calhm6, remains to be elucidated.

Receptor-operated Ca^{2+} channels: Purinergic signaling is pivotal to the physiology of microglia

in both physiological and pathological conditions⁵⁴. There are seven subtypes of ligand-gated cation channel P2X: P2X1-7. Activation of these channels in reactive microglia predominantly exerts detrimental consequences, including neuronal hyperexcitability and inflammation⁶⁸. The P2X4 subtype is particularly noted for its high calcium permeability and undergoes tight regulatory control in its translocation to the plasma membrane, with a predominant localization within lysosomal compartments. Expression of P2X4 receptors is notably increased in reactive microglial phenotypes discernible in an array of neuropathological contexts, including neuropathic pain, status epilepticus, and multiple sclerosis^{69,70}. The P2X7 receptor is another prominent Ca²⁺ entry channel in microglial cells⁷¹, which regulates IL-1 β and IL-18 release, microglial activation, and neuroinflammation⁷². The P2X7 receptor in microglial cells acts as a pattern recognition receptor (PRR) that detects extracellular ATP, a danger-associated molecular pattern (DAMP) released from cells compromised by traumatic brain injury or neurodegeneration⁷³. Despite the abundant expression of P2X7 in microglia, the channel remains inactive under normal physiological conditions, thereby limiting the intracellular ATP concentration lower than the high micromolar levels required for ion channel activation⁷⁴.

G protein-coupled receptors (GPCRs): Over 90% of non-sensory GPCRs are located in the brain. Diminished extracellular Ca²⁺ concentrations activate G protein-coupled calcium-sensing receptors, triggering the phospholipase C (PKC) signaling pathway. The presence of these receptors in microglial cells suggests their involvement in maintaining local ionic balance⁷⁵.

NCX: The NCX is a bidirectional ion transporter, facilitating the exchange of Ca²⁺ efflux and Na⁺ influx³⁸. In physiological states, the NCX primarily facilitates the influx of Na⁺ along their concentration gradient into the cellular matrix while concurrently extruding calcium ions (Ca²⁺)⁷⁶. Conversely, in pathological conditions, the NCX predominantly engages in reverse mode, which is marked by disrupted calcium homeostasis and influences a plethora of calcium-dependent processes occurring at both cellular and systemic levels⁷⁷.

MICROGLIAL ION CHANNELS IN AD

A close association between neuronal calcium dysfunction and the progression of AD (Figure 2),

characterized by the accumulation of beta-amyloid (A β) plaques, neurofibrillary tangles, and neuroinflammation^{78,79}, raises a calcium-centered hypothesis for the disease^{80,81}, which states that AD begins with early dysregulation of calcium signaling, precipitating neurodegeneration through mitochondrial dysfunction, oxidative stress, and neuroinflammation⁸². Increased basal Ca²⁺ levels, rapid depletion of Ca²⁺ from ER stores, and a reduced response to ATP stimulation are reported in microglial cells of AD patients, as compared to healthy controls⁸³.

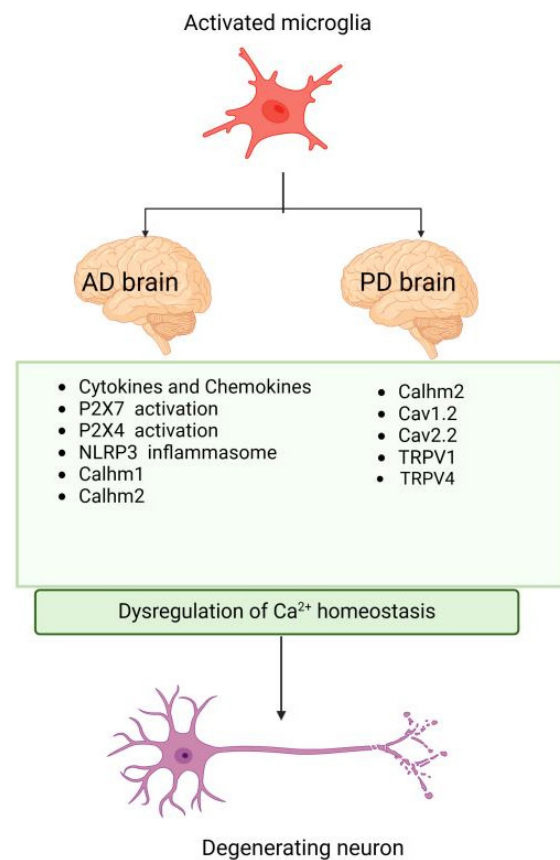


Figure 2. Schematic diagram of microglial Ca²⁺ homeostasis dysregulation in the brain of AD patients and PD patients. In AD brain: cytokines and chemokines; P2X4R, purinergic P2X4 receptor; P2X7R, purinergic P2X7 receptor; NLRP3, NOD-like receptor pyrin domain containing 3; Calhm1, 2, Calcium homeostasis modulator family member 1, 2; In PD brain: Calhm2, Calcium homeostasis Modulator family member 2; L-type VGCCs, L-type voltage-gated calcium channels, Cav1.2, Cav2.2; Transient receptor potential channel; TRPV1, TRPV4.

Numerous calcium channels and receptors (Figure 2), including Calhm2, P2X receptor, VGCCs⁸⁴, SOCE channels, and TRP channels, play a crucial role in mediating the influx of extracellular calcium^{85,86}. Calhm1 P86L single nucleotide polymorphism has been identified as a potential risk factor for AD⁵⁹. Concurrently, Calhm2 is expressed in the CNS. Our previous research shows that Calhm2 levels increase in brain samples from AD patients. Both conventional and microglial knockout of Calhm2 inhibit the inflammation, A β deposition/plaque formation, and cognitive impairments of 5 \times FAD mice. Mechanistically, Calhm2 knockout decreases the influx of extracellular calcium in microglia, reduces microglial proinflammatory activity, and enhances their ability to phagocytose A β , thereby rebalancing inflammation and phagocytosis⁸⁷.

NCX, particularly NCX3, is implicated in neuroprotection in AD, with its dysfunction linked to neuronal death via caspase-12 activation⁸⁸. Moreover, upregulation of P2X7 is observed in the brains of AD patients, particularly in microglia around amyloid plaques^{89,90}. P2X7 facilitates the release of A β -induced chemokines, while its deletion attenuates brain lesions and cognitive impairments in a transgenic amyloid mouse AD model⁸⁹. P2X7 receptor expression is increased in both human microglial cells following exposure to A β 42, and microglial cells isolated from AD patients⁹¹. Activation of the P2X7 receptor in microglia and peripheral immune cells are known to trigger the NLRP3 inflammasome, leading to caspase-1 activation and the subsequent conversion of pro-IL-1 β into its active form, IL-1 β ⁹². Disruption of NLRP3 inflammasome activation, through pharmacological or genetic knockout, inhibits A β 42-related microglial activation and neuroinflammation, enhances A β clearance through enhanced microglial phagocytosis, and substantially restores cognitive function in APP/PS1 mice⁹³. Recent research has demonstrated that activation of P2X7R promotes microglial cells migration but decreases their phagocytic capacity⁹⁴. Moreover, both pharmacological inhibition and genetic downregulation of P2X7R effectively prevent A β -induced activation of microglial cells and neuroinflammation *in vitro* and *in vivo* studies^{92,95}.

Under physiological conditions, P2X4 is expressed at a low level⁹⁶. However, under pathological conditions, P2X4 is upregulated in reactive microglia, contributing to brain-derived

neurotrophic factor (BDNF) release and the inflammatory response^{68,97}. Reactive microglia show increased P2X4 receptor expression linked to neuroinflammation, with an elevated level of P2X4 receptors in plaque-associated microglia (PAM). Current findings suggest that microglial P2X4 facilitates lysosomal ApoE degradation, thereby indirectly impacting A β clearance in AD, which may contribute to synaptic dysfunction and cognitive decline, including memory impairment. Deletion of P2X4 reverses these cognitive deficits in APP/PS1 mice, suggesting its direct role in AD-associated topographic and spatial memory changes⁹⁸.

MICROGLIAL ION CHANNELS IN PD

PD ranks as the second most prevalent neurodegenerative disease associated with aging⁹⁹. Typical symptoms of PD include bradykinesia, limb rigidity, and tremors. The key pathogenic characteristics are the gradual loss of dopaminergic (DA) neurons in the SNc and the intraneuronal inclusions, also known as Lewy bodies (LBs), which contain a significant amount of fibrillar alpha-synuclein (α -syn) aggregates^{100,101}. Age, genetics, environment, immunological condition, and sex are all risk factors for developing PD¹⁰²⁻¹⁰⁴. A typical characteristic of PD is neuroinflammation, evidenced by astrocyte and microglia activation in the brain¹⁰⁵. Moreover, disruption of calcium homeostasis is also frequently observed in models of both sporadic and familial PD¹⁰⁶.

In our recent investigation employing the 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced PD model, it was observed that both through conventional knockout and microglial-specific conditional knockout of Calhm2, markedly attenuated dopaminergic neuronal degeneration. This was accompanied by a reduction in the microglial population and a suppression of neuroinflammatory responses, culminating in a notable amelioration of motor impairments in murine models of PD¹⁰⁷.

Growing evidence demonstrates the involvement of Cav1.2 and Cav2.2 channels in the microglial cells associated with PD (Figure 2). The Cav1.2 channel, predominantly localized to the cell soma, is integral to the modulation of gene expression, whereas the Cav2.2 channel, resident at axonal terminals, is instrumental in orchestrating neurotransmitter release¹⁰⁸. Notably, both Cav1.2 and Cav2.2 channels contribute to gene expression regulation

in microglia¹⁰⁹. Furthermore, the use of specific antagonists targeting these VGCCs results in modified expression patterns of activation markers in microglial cells⁵³. Cav1.2 channels in microglia possess neuroprotective properties and could be crucial in mitigating neurodegenerative disorders in PD¹¹⁰. The Cav1.2 channel has been reported to function as an inhibitory modulator of M1 activation and a facilitative regulator for M2 activation¹¹¹. Blocking the Cav1.2 channel in microglia cells in an MPTP-induced PD mice model aggravates neurodegeneration, extends M1-type cytokine release, and exacerbates behavioral deficits, worsening PD symptoms¹¹². However, inhibiting Cav2.2 channels in microglia ameliorates symptoms of age-related brain inflammation¹⁰⁹. In the MPTP-induced PD mice model, the blockade of microglial Cav2.2 channels markedly diminished microglial accumulation at the SNc, and notably ameliorated functional deficits, thereby suggesting the neuroprotective role of microglia in PD¹¹², and that Cav2.2 channel and Cav1.2 channel have complementary functions in microglia. These results indicate that Cav1.2 and Cav2.2 channels might act as regulation in the transition between the microglial M1 and M2, providing potential targeting for the treatment of chronic inflammatory disorders.

The TRPV1 channel, permeable to Ca²⁺, is a temperature-sensitive cationic channels^{113,114}. Recent studies have revealed that conjugated complexes comprised of TRPV1 antibody (anti-TRPV1) and ultra-small Cu₂-xSe nanoparticles are capable of stimulating the TRPV1 channel on microglial cells. This activation induces a Ca²⁺ influx, subsequently triggering the ATG5 protein and the Ca²⁺/CaMKK2/AMPK/mTOR signaling cascade. Such processes facilitate microglial autophagy to phagocytose, degradation of α -synuclein, and lead to the improvement of their athletic ability and memory function¹¹⁵, suggesting the TRPV1 channel could act as a molecular target in the treatment of PD.

CALCIUM CHANNELS AS PROSPECTIVE PHARMACOLOGICAL TARGETS FOR NEURODEGENERATIVE DISEASES

Current clinical therapy for AD and PD only provides limited efficacy. Additionally, these treatments may precipitate psychiatric adverse effects. One of the treatment targets for AD and PD is neuroinflammation¹¹⁶. However, the activation of microglia can produce a variety of proinflammatory

cytokines. The effect of targeting a single inflammatory factor in the treatment of AD and PD is still uncertain. Therefore, innovative treatments for neurodegenerative diseases are critically needed. Recent studies have shown that alterations in calcium signaling involve many aspects of key features associated with AD and PD, suggesting that calcium homeostasis is a therapeutic target¹¹⁷. Calcium antagonists, notably L-type Ca²⁺ channel inhibitors, are used clinically in the management of cardiovascular pathologies, including hypertension and cardiac arrhythmias, functioning through the inhibition of voltage-dependent calcium channels^{118,119}. Moreover, calcium channel blockers might also reduce the risk and improve cognition in AD and PD.

L-type calcium channel blockers, such as Verapamil and Nimodipine, have been reported to elicit anti-inflammatory responses in microglial cells and provide neuroprotective effects¹²⁰. Verapamil not only inhibits calcium influx into neurons but also targets brain microglia, attenuating inflammation and protecting dopaminergic neurons from damage. Verapamil exhibits neuroprotective properties, not only through blocking neuronal L-type calcium channels but also by inhibiting the excessive production of microglial activation. Moreover, Nicardipine has been documented to modulate calcium signaling within glial cells^{121,122}. Studies have shown that the calcium channel blocker Nicardipine can reduce pro-inflammatory transcription factor activation and inhibit microglial activation, providing it a potential therapeutic agent for inflammation-associated neurodegenerative diseases¹²³.

Recent studies have shown that the downregulation of intracellular calcium concentrations, achieved through the targeted inhibition of calcium channel proteins, facilitates a decrease in the levels of neuroinflammation¹²⁴. Our previous study demonstrates that the conditional knockout of Calhm2 in microglia markedly reduces A β deposition/plaque formation, concurrently enhancing cognitive capabilities in the AD murine model. Similarly, knockout of Calhm2 also inhibits neuroinflammation, and rescues the decrease of tyrosine hydroxylase (TH)-positive neurons in mouse PD models induced by the MPTP, thereby improving mechanical defects in Parkinson's disease model mice, which implies that Calhm2 is instrumental in the microglial modulation of neuroinflammation¹⁰⁷. This finding suggests that targeting

microglial Calhm2 could represent a promising strategy for the new therapeutic interventions for neurodegenerative diseases¹²⁵.

CONCLUSION AND FUTURE PERSPECTIVE

In conclusion, the homeostasis of intracellular calcium (Ca^{2+}) is crucial in governing neuronal physiological processes such as proliferation, maturation, electrical activity characteristics, synaptic adaptability, and cognitive functions. Dysregulation of Ca^{2+} homeostasis in cells is associated with various neuropathological states, including necrotic and apoptotic cell death, impaired autophagic mechanisms, and neuronal degeneration. This review summarizes the significant role of calcium channels in the activation of microglia and the pathogenesis of diseases associated with neuroinflammation, especially in AD and PD, and highlights the relationship among calcium homeostasis, neuroinflammation, and neurological disorders, offering novel therapeutic strategies. Therefore, exploring effective Ca^{2+} ion channel inhibitors and elucidating their role in neurological diseases will be crucial in the future.

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Author Contributions

Shasha Wang reviewed the literature, wrote the manuscript, and prepared the figures; Jinyu Zhang, Jingdan Zhang, and Ao Li reviewed the manuscript; Jinbo Cheng and Zengqiang Yuan performed a comprehensive review of the literature.

Conflicts of Interest

The authors declare no conflicts of interest. ♦

REFERENCES

- GINHOUX, F. & PRINZ, M. Origin of microglia: current concepts and past controversies. *Cold Spring Harb Perspect Biol* 7, a020537, doi:10.1101/cshperspect.a020537 (2015).
- KORIN, B. *ET AL.* High-dimensional, single-cell characterization of the brain's immune compartment. *Nat Neurosci* 20, 1300-1309, doi:10.1038/nn.4610 (2017).
- LIU, Y. U. *ET AL.* Neuronal network activity controls microglial process surveillance in awake mice via norepinephrine signaling. *Nat Neurosci* 22, 1771-1781, doi:10.1038/s41593-019-0511-3 (2019).
- PARKHURST, C. N. *ET AL.* Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell* 155, 1596-1609, doi:10.1016/j.cell.2013.11.030 (2013).
- HOFFMANN, A., KANN, O., OHLEMEYER, C., HANISCH, U. K. & KETTENMANN, H. Elevation of basal intracellular calcium as a central element in the activation of brain macrophages (microglia): suppression of receptor-evoked calcium signaling and control of release function. *J Neurosci* 23, 4410-4419, doi:10.1523/jneurosci.23-11-04410.2003 (2003).
- CHEN, Y. & COLONNA, M. Microglia in Alzheimer's disease at single-cell level. Are there common patterns in humans and mice? *J Exp Med* 218, doi:10.1084/jem.20202717 (2021).
- KEREN-SHAUL, H. *ET AL.* A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease. *Cell* 169, 1276-1290. e1217, doi:10.1016/j.cell.2017.05.018 (2017).
- YU, T. & WEIDONG, L. Differential Roles of M1 and M2 Microglia in Neurodegenerative Diseases. *Molecular Neurobiology* 53 (2016).
- FRANCO, R. & FERNÁNDEZ-SUÁREZ, D. Alternatively activated microglia and macrophages in the central nervous system. *Prog Neurobiol* 131, 65-86, doi:10.1016/j.pneurobio.2015.05.003 (2015).
- OLSON, K. E. & GENDELMAN, H. E. Immunomodulation as a neuroprotective and therapeutic strategy for Parkinson's disease. *Curr Opin Pharmacol* 26, 87-95, doi:10.1016/j.coph.2015.10.006 (2016).
- YAO, K. & ZU, H. B. Microglial polarization: novel therapeutic mechanism against Alzheimer's disease. *Inflammopharmacology* 28, 95-110, doi:10.1007/s10787-019-00613-5 (2020).
- MIZOGUCHI, Y. & MONJI, A. Microglial Intracellular Ca^{2+} Signaling in Synaptic Development and its Alterations in Neurodevelopmental Disorders. *Front Cell Neurosci* 11, 69, doi:10.3389/fncel.2017.00069 (2017).
- SHARMA, P. & PING, L. Calcium ion influx in microglial cells: physiological and therapeutic significance. *J Neurosci Res* 92, 409-423, doi:10.1002/jnr.23344 (2014).

14. SUZUKI, Y., INOUE, T. & RA, C. L-type Ca²⁺ channels: a new player in the regulation of Ca²⁺ signaling, cell activation and cell survival in immune cells. *Mol Immunol* 47, 640-648, doi:10.1016/j.molimm.2009.10.013 (2010).
15. LUO, L. *ET AL.* Ion channels and transporters in microglial function in physiology and brain diseases. *Neurochem Int* 142, 104925, doi:10.1016/j.neuint.2020.104925 (2021).
16. TOESCU, E. C. & VERKHRATSKY, A. Role of calcium in normal aging and neurodegeneration. *Aging Cell* 6, 265, doi:10.1111/j.1474-9726.2007.00299.x (2007).
17. WANG, J., HU, W. W., JIANG, Z. & FENG, M. J. Advances in treatment of neurodegenerative diseases: Perspectives for combination of stem cells with neurotrophic factors. *World J Stem Cells* 12, 323-338, doi:10.4252/wjsc.v12.i5.323 (2020).
18. SWART, T. & HURLEY, M. J. Calcium Channel Antagonists as Disease-Modifying Therapy for Parkinson's Disease: Therapeutic Rationale and Current Status. *CNS Drugs* 30, 1127-1135, doi:10.1007/s40263-016-0393-9 (2016).
19. NIRLAULA, A., SHERIDAN, J. F. & GODBOUT, J. P. Microglia Priming with Aging and Stress. *Neuropsychopharmacology* 42, 318-333, doi:10.1038/npp.2016.185 (2017).
20. NATHAN, C. & DING, A. Nonresolving inflammation. *Cell* 140, 871-882, doi:10.1016/j.cell.2010.02.029 (2010).
21. GOLDBERG, J. *ET AL.* Targeting of intracellular Ca(2+) stores as a therapeutic strategy against age-related neurotoxicities. *NPJ Aging Mech Dis* 6, 10, doi:10.1038/s41514-020-00048-1 (2020).
22. ZÜNDORF, G. & REISER, G. Calcium dysregulation and homeostasis of neural calcium in the molecular mechanisms of neurodegenerative diseases provide multiple targets for neuroprotection. *Antioxid Redox Signal* 14, 1275-1288, doi:10.1089/ars.2010.3359 (2011).
23. BERRIDGE, M. J., BOOTMAN, M. D. & RODERICK, H. L. Calcium signalling: dynamics, homeostasis and remodelling. *Nat Rev Mol Cell Biol* 4, 517-529, doi:10.1038/nrm1155 (2003).
24. CARAFOLI, E. & KREBS, J. Why Calcium? How Calcium Became the Best Communicator. *J Biol Chem* 291, 20849-20857, doi:10.1074/jbc.R116.735894 (2016).
25. CLAPHAM, D. E. Calcium signaling. *Cell* 131, 1047-1058, doi:10.1016/j.cell.2007.11.028 (2007).
26. BAGUR, R. & HAJNOCZKY, G. Intracellular Ca(2+) Sensing: Its Role in Calcium Homeostasis and Signaling. *Molecular Cell* 66, 780-788, doi:10.1016/j.molcel.2017.05.028 (2017).
27. FARBER, K. & KETTENMANN, H. Functional role of calcium signals for microglial function. *Glia* 54, 656-665, doi:10.1002/glia.20412 (2006).
28. GEHRMANN, J., MATSUMOTO, Y. & KREUTZBERG, G. W. Microglia: intrinsic immune effector cell of the brain. *Brain Res Brain Res Rev* 20, 269-287, doi:10.1016/0165-0173(94)00015-h (1995).
29. PAN, K. & GARASCHUK, O. The role of intracellular calcium-store-mediated calcium signals in in vivo sensor and effector functions of microglia. *J Physiol*, doi:10.1113/jp279521 (2022).
30. OLMEDILLAS DEL MORAL, M., ASAVAPANUMAS, N., UZCÁTEGUI, N. L. & GARASCHUK, O. Healthy Brain Aging Modifies Microglial Calcium Signaling In Vivo. *Int J Mol Sci* 20, doi:10.3390/ijms20030589 (2019).
31. KETTENMANN, H., HANISCH, U. K., NODA, M. & VERKHRATSKY, A. Physiology of microglia. *Physiol Rev* 91, 461-553, doi:10.1152/physrev.00011.2010 (2011).
32. PCHITSKAYA, E., POPUGAEVA, E. & BEZPROZVANY, I. Calcium signaling and molecular mechanisms underlying neurodegenerative diseases. *Cell Calcium* 70, 87-94, doi:10.1016/j.ceca.2017.06.008 (2018).
33. STEPHENSON, J., NUTMA, E., VAN DER VALK, P. & AMOR, S. Inflammation in CNS neurodegenerative diseases. *Immunology* 154, 204-219, doi:10.1111/imm.12922 (2018).
34. IACOPINO, A. M. & CHRISTAKOS, S. Specific reduction of calcium-binding protein (28-kilodalton calbindin-D) gene expression in aging and neurodegenerative diseases. *Proc Natl Acad Sci U S A* 87, 4078-4082, doi:10.1073/pnas.87.11.4078 (1990).
35. MATTSON, M. P., RYCHLIK, B., CHU, C. & CHRISTAKOS, S. Evidence for calcium-reducing and excitoprotective roles for the calcium-binding protein calbindin-D28k in cultured hippocampal neurons. *Neuron* 6, 41-51, doi:10.1016/0896-6273(91)90120-o (1991).
36. MATTSON, M. P., GLEICHMANN, M. & CHENG, A. Mitochondria in neuroplasticity and neurological disorders. *Neuron* 60, 748-766, doi:10.1016/j.neuron.2008.10.010 (2008).
37. MICHAELIS, M., NIESWANDT, B., STEGNER, D., EILERS, J. & KRAFT, R. STIM1, STIM2, and Orai1 regulate store-operated calcium entry and purinergic activation of microglia. *Glia* 63, 652-663, doi:10.1002/glia.22775 (2015).

38. GILADI, M., SHOR, R., LISNYANSKY, M. & KHAN-ANSHVILI, D. Structure-Functional Basis of Ion Transport in Sodium-Calcium Exchanger (NCX) Proteins. *Int J Mol Sci* 17, doi:10.3390/ijms17111949 (2016).
39. HEO, D. K., LIM, H. M., NAM, J. H., LEE, M. G. & KIM, J. Y. Regulation of phagocytosis and cytokine secretion by store-operated calcium entry in primary isolated murine microglia. *Cell Signal* 27, 177-186, doi:10.1016/j.cell-sig.2014.11.003 (2015).
40. LIM, D. ET AL. Calcium signalling toolkits in astrocytes and spatio-temporal progression of Alzheimer's disease. *Curr Alzheimer Res* 13, 359-369, doi:10.2174/1567205013666151116130104 (2016).
41. ZHANG, E. & LIAO, P. Brain transient receptor potential channels and stroke. *J Neurosci Res* 93, 11651183, doi:10.1002/jnr.23529 (2015).
42. NILIUS, B., OWSIANIK, G., VOETS, T. & PETERS, J. A. Transient receptor potential cation channels in disease. *Physiol Rev* 87, 165-217, doi:10.1152/physrev.00021.2006 (2007).
43. ECHEVERRY, S., RODRIGUEZ, M. J. & TORRES, Y. P. Transient Receptor Potential Channels in Microglia: Roles in Physiology and Disease. *Neurotox Res* 30, 467-478, doi:10.1007/s12640-016-9632-6 (2016).
44. SHIRAKAWA, H. & KANEKO, S. Physiological and Pathophysiological Roles of Transient Receptor Potential Channels in Microglia-Related CNS Inflammatory Diseases. *Biol Pharm Bull* 41, 1152-1157, doi:10.1248/bpb.b18-00319 (2018).
45. PEDERSEN, S. F., OWSIANIK, G. & NILIUS, B. TRP channels: an overview. *Cell Calcium* 38, 233-252, doi:10.1016/j.ceca.2005.06.028 (2005).
46. MIYAKE, T., SHIRAKAWA, H., NAKAGAWA, T. & KANEKO, S. Activation of mitochondrial transient receptor potential vanilloid 1 channel contributes to microglial migration. *Glia* 63, 1870-1882, doi:10.1002/glia.22854 (2015).
47. CATTERALL, W. A., PEREZ-REYES, E., SNUTCH, T. P. & STRIESSNIG, J. International Union of Pharmacology. XLVIII. Nomenclature and structure-function relationships of voltage-gated calcium channels. *Pharmacol Rev* 57, 411-425, doi:10.1124/pr.57.4.5 (2005).
48. ERTEL, E. A. ET AL. Nomenclature of voltage-gated calcium channels. *Neuron* 25, 533-535, doi:10.1016/s0896-6273(00)81057-0 (2000).
49. TANABE, T., BEAM, K. G., ADAMS, B. A., NIDOME, T. & NUMA, S. Regions of the skeletal muscle dihydropyridine receptor critical for excitation-contraction coupling. *Nature* 346, 567-569, doi:10.1038/346567a0 (1990).
50. TANABE, T., BEAM, K. G., POWELL, J. A. & NUMA, S. Restoration of excitation-contraction coupling and slow calcium current in dysgenic muscle by dihydropyridine receptor complementary DNA. *Nature* 336, 134-139, doi:10.1038/336134a0 (1988).
51. TANABE, T., MIKAMI, A., NUMA, S. & BEAM, K. G. Cardiac-type excitation-contraction coupling in dysgenic skeletal muscle injected with cardiac dihydropyridine receptor cDNA. *Nature* 344, 451-453, doi:10.1038/344451a0 (1990).
52. TANABE, T. ET AL. Primary structure of the receptor for calcium channel blockers from skeletal muscle. *Nature* 328, 313-318, doi:10.1038/328313a0 (1987).
53. WANG, X., SAEGUSA, H., HUNTULA, S. & TANABE, T. Blockade of microglial Cav1.2 Ca(2+) channel exacerbates the symptoms in a Parkinson's disease model. *Sci Rep* 9, 9138, doi:10.1038/s41598-019-45681-3 (2019).
54. NANOU, E. & CATTERALL, W. A. Calcium Channels, Synaptic Plasticity, and Neuropsychiatric Disease. *Neuron* 98, 466-481, doi:10.1016/j.neuron.2018.03.017 (2018).
55. LISS, B. & STRIESSNIG, J. The Potential of L-Type Calcium Channels as a Drug Target for Neuroprotective Therapy in Parkinson's Disease. *Annu Rev Pharmacol Toxicol* 59, 263-289, doi:10.1146/annurev-pharmtox-010818-021214 (2019).
56. GUZMAN, J. N. ET AL. Systemic isradipine treatment diminishes calcium-dependent mitochondrial oxidant stress. *J Clin Invest* 128, 2266-2280, doi:10.1172/jci95898 (2018).
57. CATTERALL, W. A. Voltage-gated calcium channels. *Cold Spring Harb Perspect Biol* 3, a003947, doi:10.1101/cshperspect.a003947 (2011).
58. NEUMAIER, F., DIBUÉ-ADJEI, M., HESCHELER, J. & SCHNEIDER, T. Voltage-gated calcium channels: Determinants of channel function and modulation by inorganic cations. *Prog Neurobiol* 129, 1-36, doi:10.1016/j.pneurobio.2014.12.003 (2015).
59. DRESES-WERRINGLOER, U. ET AL. A polymorphism in CALHM1 influences Ca²⁺ homeostasis, Aβ levels, and Alzheimer's disease risk. *Cell* 133, 1149-1161, doi:10.1016/j.cell.2008.05.048 (2008).
60. LAMBERT, J. C. ET AL. The CALHM1 P86L polymorphism is a genetic modifier of age at onset in Alzheimer's disease: a meta-analysis study. *J Alzheimers Dis* 22, 247-255, doi:10.3233/jad-2010-100933 (2010).

61. TARUNO, A. *ET AL.* CALHM1 ion channel mediates purinergic neurotransmission of sweet, bitter and umami tastes. *Nature* 495, 223-226, doi:10.1038/nature11906 (2013).
62. MA, Z. *ET AL.* Calcium homeostasis modulator 1 (CALHM1) is the pore-forming subunit of an ion channel that mediates extracellular Ca²⁺ regulation of neuronal excitability. *Proc Natl Acad Sci U S A* 109, E1963-1971, doi:10.1073/pnas.1204023109 (2012).
63. WU, J. *ET AL.* Generation of Calhm1 knockout mouse and characterization of calhm1 gene expression. *Protein Cell* 3, 470-480, doi:10.1007/s13238-012-2932-6 (2012).
64. MA, Z. *ET AL.* CALHM3 Is Essential for Rapid Ion Channel-Mediated Purinergic Neurotransmission of GPCR-Mediated Tastes. *Neuron* 98, 547-561. e510, doi:10.1016/j.neuron.2018.03.043 (2018).
65. CHOI, W., CLEMENTE, N., SUN, W., DU, J. & LÜ, W. The structures and gating mechanism of human calcium homeostasis modulator 2. *Nature* 576, 163-167, doi:10.1038/s41586-019-1781-3 (2019).
66. SYRJANEN, J. L. *ET AL.* Structure and assembly of calcium homeostasis modulator proteins. *Nat Struct Mol Biol* 27, 150-159, doi:10.1038/s41594-019-0369-9 (2020).
67. MA, J. *ET AL.* Calhm2 governs astrocytic ATP releasing in the development of depression-like behaviors. *Mol Psychiatry* 23, 883-891, doi:10.1038/mp.2017.229 (2018).
68. MONTILLA, A., MATA, G. P., MATUTE, C. & DOMERCO, M. Contribution of P2X4 Receptors to CNS Function and Pathophysiology. *Int J Mol Sci* 21, doi:10.3390/ijms21155562 (2020).
69. FÄRBER, K. & KETTENMANN, H. Purinergic signaling and microglia. *Pflugers Arch* 452, 615-621, doi:10.1007/s00424-006-0064-7 (2006).
70. RASSENDREN, F. & AUDINAT, E. Purinergic signaling in epilepsy. *J Neurosci Res* 94, 781-793, doi:10.1002/jnr.23770 (2016).
71. HE, Y., TAYLOR, N., FOURGEAUD, L. & BHATTACHARYA, A. The role of microglial P2X7: modulation of cell death and cytokine release. *J Neuroinflammation* 14, 135, doi:10.1186/s12974-017-0904-8 (2017).
72. BHATTACHARYA, A. & BIBER, K. The microglial ATP-gated ion channel P2X7 as a CNS drug target. *Glia* 64, 1772-1787, doi:10.1002/glia.23001 (2016).
73. JANKOWSKY, J. L. *ET AL.* Mutant presenilins specifically elevate the levels of the 42 residue beta-amyloid peptide in vivo: evidence for augmentation of a 42-specific gamma secretase. *Hum Mol Genet* 13, 159-170, doi:10.1093/hmg/ddh019 (2004).
74. MONIF, M., BURNSTOCK, G. & WILLIAMS, D. A. Microglia: proliferation and activation driven by the P2X7 receptor. *Int J Biochem Cell Biol* 42, 1753-1756, doi:10.1016/j.biocel.2010.06.021 (2010).
75. BREITWIESER, G. E. Extracellular calcium as an integrator of tissue function. *Int J Biochem Cell Biol* 40, 1467-1480, doi:10.1016/j.biocel.2008.01.019 (2008).
76. TAKUMA, K., AGO, Y. & MATSUDA, T. The glial sodium-calcium exchanger: a new target for nitric oxide-mediated cellular toxicity. *Curr Protein Pept Sci* 14, 43-50, doi:10.2174/1389203711314010007 (2013).
77. TÓTH, N. *ET AL.* The reverse mode of the Na⁽⁺⁾/Ca⁽²⁺⁾ exchanger contributes to the pacemaker mechanism in rabbit sinus node cells. *Sci Rep* 12, 21830, doi:10.1038/s41598-022-25574-8 (2022).
78. 2012 Alzheimer's disease facts and figures. *Alzheimers Dement* 8, 131-168, doi:10.1016/j.jalz.2012.02.001 (2012).
79. TIRABOSCHI, P., HANSEN, L. A., THAL, L. J. & COREY-BLOOM, J. The importance of neuritic plaques and tangles to the development and evolution of AD. *Neurology* 62, 1984-1989, doi:10.1212/01.wnl.0000129697.01779.0a (2004).
80. Calcium Hypothesis of Alzheimer's disease and brain aging: A framework for integrating new evidence into a comprehensive theory of pathogenesis. *Alzheimers Dement* 13, 178-182.e117, doi:10.1016/j.jalz.2016.12.006 (2017).
81. KHACHATURIAN, Z. S. Calcium, membranes, aging, and Alzheimer's disease. Introduction and overview. *Ann N Y Acad Sci* 568, 1-4, doi:10.1111/j.1749-6632.1989.tb12485.x (1989).
82. CUMMINGS, J. L., TONG, G. & BALLARD, C. Treatment Combinations for Alzheimer's Disease: Current and Future Pharmacotherapy Options. *J Alzheimers Dis* 67, 779-794, doi:10.3233/jad-180766 (2019).
83. MCLARNON, J. G., CHOI, H. B., LUE, L. F., WALKER, D. G. & KIM, S. U. Perturbations in calcium-mediated signal transduction in microglia from Alzheimer's disease patients. *J Neurosci Res* 81, 426-435, doi:10.1002/jnr.20487 (2005).
84. ANEKONDA, T. S. *ET AL.* L-type voltage-gated calcium channel blockade with isradipine as a therapeutic strategy for Alzheimer's disease. *Neurobiol Dis* 41, 62-70, doi:10.1016/j.nbd.2010.08.020 (2011).

85. GOODISON, W. V., FRISARDI, V. & KEHOE, P. G. Calcium channel blockers and Alzheimer's disease: potential relevance in treatment strategies of metabolic syndrome. *J Alzheimers Dis* 30 Suppl 2, S269-282, doi:10.3233/jad-2012-111664 (2012).
86. MORENO-ORTEGA, A. J. ET AL. CALHM1 and its polymorphism P86L differentially control Ca^{2+} homeostasis, mitogen-activated protein kinase signaling, and cell vulnerability upon exposure to amyloid β . *Aging Cell* 14, 1094-1102, doi:10.1111/ace1.12403 (2015).
87. CHENG, J. ET AL. Microglial Calhm2 regulates neuroinflammation and contributes to Alzheimer's disease pathology. *Sci Adv* 7, doi:10.1126/sciadv.abe3600 (2021).
88. PICCIALLI, I. ET AL. The Na^{+}/Ca^{2+} Exchanger 3 Is Functionally Coupled With the Na^{+} Voltage-Gated Channel and Promotes an Endoplasmic Reticulum Ca^{2+} Refilling in a Transgenic Model of Alzheimer's Disease. *Front Pharmacol* 12, 775271, doi:10.3389/fphar.2021.775271 (2021).
89. MARTIN, E. ET AL. New role of P2X7 receptor in an Alzheimer's disease mouse model. *Mol Psychiatry* 24, 108-125, doi:10.1038/s41380-018-0108-3 (2019).
90. MCLARNON, J. G., RYU, J. K., WALKER, D. G. & CHOI, H. B. Upregulated expression of purinergic P2X(7) receptor in Alzheimer disease and amyloid-beta peptide-treated microglia and in peptide-injected rat hippocampus. *J Neuro-pathol Exp Neurol* 65, 1090-1097, doi:10.1097/01.jnen.0000240470.97295.d3 (2006).
91. RONNING, K. E. ET AL. The P2X7 Receptor, a Multifaceted Receptor in Alzheimer's Disease. *Int J Mol Sci* 24, doi:10.3390/ijms241411747 (2023).
92. SANZ, J. M. ET AL. Activation of microglia by amyloid β requires P2X7 receptor expression. *J Immunol* 182, 4378-4385, doi:10.4049/jimmunol.0803612 (2009).
93. HENEKA, M. T., MCMANUS, R. M. & LATZ, E. Inflammasome signalling in brain function and neurodegenerative disease. *Nat Rev Neurosci* 19, 610-621, doi:10.1038/s41583-018-0055-7 (2018).
94. MARTÍNEZ-FRAILES, C. ET AL. Amyloid Peptide Induced Neuroinflammation Increases the P2X7 Receptor Expression in Microglial Cells, Impacting on Its Functionality. *Front Cell Neurosci* 13, 143, doi:10.3389/fncel.2019.00143 (2019).
95. CHEN, X. ET AL. Brilliant Blue G improves cognition in an animal model of Alzheimer's disease and inhibits amyloid- β -induced loss of filopodia and dendrite spines in hippocampal neurons. *Neuroscience* 279, 94-101, doi:10.1016/j.neuroscience.2014.08.036 (2014).
96. XU, J. ET AL. P2X4 Receptor Reporter Mice: Sparse Brain Expression and Feeding-Related Presynaptic Facilitation in the Arcuate Nucleus. *J Neurosci* 36, 8902-8920, doi:10.1523/jneurosci.1496-16.2016 (2016).
97. ULMANN, L. ET AL. Involvement of P2X4 receptors in hippocampal microglial activation after status epilepticus. *Glia* 61, 1306-1319, doi:10.1002/glia.22516 (2013).
98. HUA, J. ET AL. Microglial P2X4 receptors promote ApoE degradation and contribute to memory deficits in Alzheimer's disease. *Cell Mol Life Sci* 80, 138, doi:10.1007/s00018-023-04784-x (2023).
99. DE LAU, L. M. & BRETHER, M. M. Epidemiology of Parkinson's disease. *Lancet Neurol* 5, 525-535, doi:10.1016/s1474-4422(06)70471-9 (2006).
100. MOORE, D. J., WEST, A. B., DAWSON, V. L. & DAWSON, T. M. Molecular pathophysiology of Parkinson's disease. *Annu Rev Neurosci* 28, 57-87, doi:10.1146/annurev.neuro.28.061604.135718 (2005).
101. PISTACCHI, M. ET AL. Gait analysis and clinical correlations in early Parkinson's disease. *Funct Neurol* 32, 28-34, doi:10.11138/fneur/2017.32.1.028 (2017).
102. CERRI, S., MUS, L. & BLANDINI, F. Parkinson's Disease in Women and Men: What's the Difference? *J Parkinsons Dis* 9, 501-515, doi:10.3233/jpd-191683 (2019).
103. PAJARES, M., A, I. R., MANDA, G., BOSCA, L. & CUADRADO, A. Inflammation in Parkinson's Disease: Mechanisms and Therapeutic Implications. *Cells* 9, doi:10.3390/cells9071687 (2020).
104. TANSEY, M. G. ET AL. Inflammation and immune dysfunction in Parkinson disease. *Nat Rev Immunol* 22, 657-673, doi:10.1038/s41577-022-00684-6 (2022).
105. KWON, H. S. & KOH, S. H. Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. *Transl Neurodegener* 9, 42, doi:10.1186/s40035-020-00221-2 (2020).
106. POST, M. R., LIEBERMAN, O. J. & MOSHAROV, E. V. Can Interactions Between α -Synuclein, Dopamine and Calcium Explain Selective Neurodegeneration in Parkinson's Disease? *Front Neurosci* 12, 161, doi:10.3389/fnins.2018.00161 (2018).

107. BO, X. *ET AL.* Deletion of Calhm2 alleviates MPTP-induced Parkinson's disease pathology by inhibiting EFHD2-STAT3 signaling in microglia. *Theranostics* 13, 1809-1822, doi:10.7150/thno.83082 (2023).
108. LIN, Y., McDONOUGH, S. I. & LIPSCOMBE, D. Alternative splicing in the voltage-sensing region of N-Type CaV2.2 channels modulates channel kinetics. *J Neurophysiol* 92, 2820-2830, doi:10.1152/jn.00048.2004 (2004).
109. HUNTULA, S., SAEGUSA, H., WANG, X., ZONG, S. & TANABE, T. Involvement of N-type Ca(2+) channel in microglial activation and its implications to aging-induced exaggerated cytokine response. *Cell Calcium* 82, 102059, doi:10.1016/j.ceca.2019.102059 (2019).
110. MADRY, C. *ET AL.* Microglial Ramification, Surveillance, and Interleukin-1 β Release Are Regulated by the Two-Pore Domain K(+) Channel THIK-1. *Neuron* 97, 299-312.e296, doi:10.1016/j.neuron.2017.12.002 (2018).
111. ROUX, B. Ion channels and ion selectivity. *Essays Biochem* 61, 201-209, doi:10.1042/ebc20160074 (2017).
112. SAEGUSA, H., LI, X., WANG, X., KAYAKIRI, M. & TANABE, T. Knockdown of microglial Cav2.2 N-type voltage-dependent Ca(2+) channel ameliorates behavioral deficits in a mouse model of Parkinson's disease. *FEBS Lett* 594, 2914-2922, doi:10.1002/1873-3468.13853 (2020).
113. CATERINA, M. J. *ET AL.* The capsaicin receptor: a heat-activated ion channel in the pain pathway. *Nature* 389, 816-824, doi:10.1038/39807 (1997).
114. FARFARIELLO, V., AMANTINI, C. & SANTONI, G. Transient receptor potential vanilloid 1 activation induces autophagy in thymocytes through ROS-regulated AMPK and Atg4C pathways. *J Leukoc Biol* 92, 421-431, doi:10.1189/jlb.0312123 (2012).
115. YUAN, J. *ET AL.* Controlled Activation of TRPV1 Channels on Microglia to Boost Their Autophagy for Clearance of Alpha-Synuclein and Enhance Therapy of Parkinson's Disease. *Adv Mater* 34, e2108435, doi:10.1002/adma.202108435 (2022).
116. ATHAUDA, D. & FOLTYNIE, T. The ongoing pursuit of neuroprotective therapies in Parkinson disease. *Nat Rev Neurol* 11, 25-40, doi:10.1038/nrneurol.2014.226 (2015).
117. SCHRANK, S., BARRINGTON, N. & STUTZMANN, G. E. Calcium-Handling Defects and Neurodegenerative Disease. *Cold Spring Harb Perspect Biol* 12, doi:10.1101/cshperspect.a035212 (2020).
118. FLECKENSTEIN, A. History of calcium antagonists. *Circ Res* 52, I3-16 (1983).
119. McDONOUGH, S. I. Gating modifier toxins of voltage-gated calcium channels. *Toxicon* 49, 202-212, doi:10.1016/j.toxicon.2006.09.018 (2007).
120. MICHELUCCI, A., HEURTAUX, T., GRANDBARBE, L., MORGA, E. & HEUSCHLING, P. Characterization of the microglial phenotype under specific pro-inflammatory and anti-inflammatory conditions: Effects of oligomeric and fibrillar amyloid-beta. *J Neuroimmunol* 210, 3-12, doi:10.1016/j.jneuroim.2009.02.003 (2009).
121. KOIZUMI, S. *ET AL.* Spatial and temporal aspects of Ca²⁺ signaling mediated by P2Y receptors in cultured rat hippocampal astrocytes. *Life Sci* 72, 431-442, doi:10.1016/s0024-3205(02)02273-7 (2002).
122. KOMAGIRI, Y., NAKAMURA, K. & KUBOKAWA, M. A nifedipine-sensitive Ca²⁺ entry contributes to the hypotonicity-induced increase in [Ca²⁺]_i of principal cells in rat cortical collecting duct. *Cell Calcium* 49, 35-42, doi:10.1016/j.ceca.2010.11.006 (2011).
123. HUANG, B. R. *ET AL.* Anti-neuroinflammatory effects of the calcium channel blocker nifedipine on microglial cells: implications for neuroprotection. *PLoS One* 9, e91167, doi:10.1371/journal.pone.0091167 (2014).
124. PARK, J. H. *ET AL.* Lomerizine inhibits LPS-mediated neuroinflammation and tau hyperphosphorylation by modulating NLRP3, DYRK1A, and GSK3 α/β . *Front Immunol* 14, 1150940, doi:10.3389/fimmu.2023.1150940 (2023).
125. DONG, Y. & TANG, L. Microglial Calcium Homeostasis Modulator 2: Novel Anti-neuroinflammation Target for the Treatment of Neurodegenerative Diseases. *Neurosci Bull*, doi:10.1007/s12264-023-01153-3 (2023).





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