

Blood biomarkers in MCI conversion to Alzheimer's disease: a systematic review and meta-analysis

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ABSTRACT

Background: The predictive effects of blood biomarkers (BBMs) in the progression of Alzheimer's disease (AD) have been reported recently. However, controversies still exist. In the present study, we aim to identify the predictive performances of BBMs in the conversion from Mild cognitive impairment (MCI) to AD.

Methods: PubMed, Embase, Cochrane Library, and Web of Science from inception to June 10, 2023 were searched. Predictive potentials were evaluated by pooling the ratio of means (ROMs), relative risks (RRs), and diagnostic indexes from MCI-converters (MCI-c: MCI patients who convert to AD) and MCI-non converters (MCI-nc) based on fixed-effects or random-effects. Newcastle-Ottawa Quality Assessment Scale (NOS) was applied for quality assessment.

Results: A total of 44 studies with 9343 participants from 28 cohorts were included in the meta-analysis, whereas the other 45 articles were included in the qualitative review. The average score of 44 studies included in the meta-analysis was 7.125. In pooled ROMs, plasma A\u03c342/A\u03c340 was lower, whereas AB40, T-tau, P-tau 181, P-tau 217, NFL, and GFAP were higher in MCI-c than MCI-nc. In pooled RRs, P-tau (RR=2.50, 95%CI: 2.04-3.06) as a continuous variable, AB42/AB40 as a categorical variable (RR=1.28, 95%Cl: 1.01-1.61) could predict future conversion risk of MCI patients. In diagnostic indexes, the diagnostic odds ratio (DOR) was 42 for P-tau 217 (sensitivity: 91%; specificity: 81%), 15 for P-tau 181 (sensitivity: 81%; specificity: 78%), 12.71 for GFAP (sensitivity: 71%; specificity: 86%), 6 for AB42/AB40 (sensitivity: 86%; specificity: 49%, and 6 for NFL) (sensitivity: 80%; specificity: 61%).

Conclusion: Here, our results indicated that blood biomarkers held promising potential in predicting MCI conversion. However, more prospective cohorts based on particular MCI types and high-sensitivity assays are warranted to validate the results next.

KEYWORDS: blood biomarkers; mild cognitive impairment; Alzheimer's disease.

1. INTRODUCTION

As the most common type of dementia worldwide, Alzheimer's disease (AD) is a neurodegenerative disorder characterized by cognitive decline and progressive memory loss. With the advancing aging of the global population, about 1 in 9 people over age 65 have AD,

two-thirds of which are women. Mortality due to AD between 2000 and 2019 has more than doubled, increasing by 145%, resulting in an enormous disease burden ("2023 Alzheimer's disease facts and figures," 2023; J. T. Wang et al., 2022). Therefore, early identification and intervention play a fundamental role in AD.

The evolution of the diagnostic criteria for AD epitomizes the convergence of clinical observation with advanced biotechnological insights. Before 1984, the standardized criteria for AD diagnoses were inconsistent. The introduction of the National Institute of Neurological and Communicative Disorders and Stroke - Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA) in 1984 marked a propounding shift, delineating "possible," "probable," and "definite" AD diagnostic categories (G. McKhann et al., 1984). After 2000, advancements in neuroimaging and CSF analyses highlighted biomarkers like amyloid plaques and tau tangles, enhancing diagnostic precision. The 2011 NIA-AA criteria integrated these biomarkers, underscoring "preclinical AD" stages (G. M. McKhann et al., 2011). The 2018 NIA-AA revision pioneered the AT(N) framework that consisted of AB deposition, tau pathology, and neurodegeneration (Jack et al., 2018). More recently, the introduction and renovation of biomarkers of ATNIVS frame in 2023 NIA-AA (A: Aβ proteinopathy; T: AD tau proteinopathy; N: injury, dysfunction, or degeneration of neuropil; I: inflammation, astrocytic activation; V: vascular brain injury; S: α-synuclein) incorporated blood biomarkers for AD diagnosis and staging. This underpinned current AD diagnosis, promoting accurate detection and refining research avenues.

Generally speaking, the AD continuum comprises preclinical AD, mild cognitive impairment (MCI) due to AD, and AD dementia. Of note, heterogeneous MCI can be divided into 2 clinical phenotypes, amnestic MCI (aMCI) and non-amnestic MCI (naMCI) (Petersen, 2004), based on the presence of memory impairment. aMCI is regarded as prodromal AD, while naMCI tends to develop non-AD dementia. When incorporating biomarkers, MCI can also be diagnosed as MCI due to AD with positive pathophysiological biomarkers (Albert et al., 2011). Among MCI patients, around 15% develop dementia after two years; one-third progress to AD dementia within five years. However, 26% MCI patients will reverse cognitive normal. Hence, finding reliable biomarkers and identifying MCI subjects prone to develop dementia is a crucial task of current research ("2023 Alzheimer's disease facts and figures," 2023).

However, to some extent, expensive PET imaging and invasive lumber puncture have restricted the application of imaging CSF examinations. Here, blood-based biomarkers (BBM) might be a resourceful tool for appraising AD risk and tailoring interventions with their cost-effective, less invasive, and serially-measured nature. First and foremost, plasma Aβ and P-tau are associated with corresponding levels in CSF, with Aβ-PET (Barthélemy, Horie, Sato, & Bateman, 2020) or tau-PET scans (Bilgel et al., 2023), and post-mortem AD pathology (Z. B. Wang et al., 2023), differentiating AD dementia from other forms of dementia (Hampel et al., 2021; Hansson et al., 2022; Teunissen et al., 2022). Secondly, in temporal order, plasma P-tau 217 and P-tau 231 can capture the earliest brain amyloid alterations before the overt amyloid plaque is formed. Besides, Aβ42/ Aβ40 declines 41 years before Aβ-PET positivity. Thirdly, accumulating evidence has reported the predictive roles of blood biomarkers in AD continuum. Taken together, the aforementioned aspects highlight BBMs as surrogate biomarkers in the progression of AD.

Nevertheless, the predictive effects of candidate BBMs in the conversion from MCI to AD remain undetermined so far. Hence, we conducted a comprehensive meta-analysis focusing mainly on four categories of biomarkers—amyloid, phosphorylated tau (P-tau), neurofilament light (NFL), and glial fibrillary acidic protein (GFAP). Meanwhile, to optimize the generalizability and scalability of the pooled results, we discussed the variances between MCI and aMCI groups given that aMCI patients are more prone to develop AD compared with naMCI and that different diagnostic might bring heterogeneity of these results. Besides, we also deliberate on discrepancies among different measuring methods. In the end, we also summarized other novel and emerging biomarkers in the qualitative review.

2. MATERIALS AND METHODS

This meta-analysis was registered in PROSPERO (https://www.crd.york.ac.uk/PROSPERO/) with a CRD of 42023404506 and adhered to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement (McInnes et al., 2018) (Supplementary material 1).

2.1. Search strategy

PubMed, Web of Science, Cochrane Library, and Embase from inception to June 10, 2023 were independently searched by two study investigators (HL and JW). The following three parts of search strings were applied and combined with Boolean Operator "AND" (Supplementary material 2). The first section referred to AD and MCI. The second referred to blood biomarkers, and the third to study types. To avoid omission, we manually scrutinized the references of key retrieved articles and relevant reviews.

2.2. Selection criteria

Articles were included if they satisfied the following four conditions: (1) articles from peer-reviewed journals and investigating human participants; (2) the diagnosis of MCI and AD are in agreement with recognized diagnostic criteria. (3) reporting at least one AD blood-based biomarker at baseline. (4) for longitudinal investigations, the clinical diagnosis of the participants should be reported in the follow-up duration. The exclusion criteria are demonstrated in **Figure 1.** To avoid double-counting participants by including more than one publication from the same cohort, we only selected one of the publications that reported the largest number of cases or had the longest follow-up duration. The literature selection was performed by two experienced investigators (HL and WJ). Discrepancies were resolved by consensus with a third neurologist (GW).

2.3. Data extraction and quality assessment

Two reviewers (HL and WJ) independently implemented data extraction and quality appraisal of included studies. Discrepancies were resolved by discussing with a third neurologist (GW). The following information was extracted: (1) basic information — the name of the first author, publication year, study cohort, region, study type, follow-up duration, sample type, assay methods; (2) subjects' information-MCI criteria, AD criteria gender, MCI type, the number of MCI converters and non-converters, APOE carriers, education years, baseline age and MMSE; (3) blood biomarker's information-baseline concentrations of blood biomarkers, effect sizes and 95% CIs, true-positive (TP), false-positive (FP), true-negative (TN), and

false-negative (FN) values. The authors were contacted for essential information. The NOS scale was exploited for quality assessment (Stang, 2010). Studies scoring 7–9 are of high quality, 4–6 of moderate quality, and less than 4 of low quality.

2.4. Statistical analysis

Blood biomarkers reported in no less than four studies were pooled in meta-analysis. The primary outcome was the clinical conversion from MCI to AD, further denoted by ratios of means (ROM), relative risks (RR), and diagnostic indicators. ROM method was used to calculate the ratio of biomarker concentrations between MCI-c and MCI-nc. A ratio above one indicates that the level of BBM is higher in MCI-c than that of MCI-nc (Friedrich, Adhikari, & Beyene, 2008; Olsson et al., 2016). Heterogeneity was examined by Q test and I2 statistic. I2 values >50% or p<0.1 of Q test represented significant heterogeneity. A fixed-effects model was applied when p>0.1 and $I^2 <50\%$; otherwise, a random-effects model was used. To investigate the sources of heterogeneity and respective pooled effects among different groups, subgroup analyses were conducted based on MCI type, methods, region, study type, follow-up duration, baseline age, female percent, APOE $\varepsilon 4$ status, and MMSE. Meta-regression analyses were also applied. Sensitivity analyses were employed by a leave-one-out strategy to detect the stability of the pooled results. Publication bias was assessed by Egger's tests and visualized by the funnel plots. "Trim and fill" strategy was used to assess whether pooled results were affected by publication bias. For diagnostic indexes, the pooled sensitivity, specificity, PLR, NLR, and DOR were computed with TP, FP, TN, and FN. HSROC curve was adopted to evaluate the correlation between sensitivity and specificity. Deek's funnel plot asymmetry test was employed to test publication bias. All statistical analyses were performed using STATA version 15. P-values less than 0.05 were considered significant. Biomarkers that were reported in less than 4 studies were mainly discussed in the qualitative review.

3. RESULTS

3.1. Characteristics of included studies and quality appraisal

After retrieving 6718 studies, there were 89 eligible studies (Supplementary material 3). Forty-four

studies containing 10 biomarkers were included in meta-analyses, while the other 45 articles were included in systematic review due to the insufficient number of studies (<4) (Figure 1). In all, 44 studies enrolled a total of 9343 participants from 28 cohorts with sample sizes ranging from 33 to 584 subjects. The characteristics of the included studies are listed in **Table 1.** Twenty-eight studies reported baseline concentrations of blood biomarkers (Supplementary material 4); Twenty-two studies measured effect sizes and 95%Cis (Supplementary material 4); Twelve studies reported diagnostic indexes (Supplementary material 4). Overall, 10 blood biomarkers (Aβ42, Aβ40, Aβ42/ Aβ40, T-tau, P-tau 181, P-tau 217, NFL, GFAP, BDNF, and APOE) were identified. The average NOS score of the included studies was 7.125, all of which were rated as having moderate to high quality (Supplemental material 5).

3.2. Results of meta-analysis

3.2.1. ROMs of blood biomarkers

Aβ42, Aβ40, Aβ42/Aβ40, T-tau, P-tau 181, P-tau 217, NFL, GFAP, and BDNF were analyzed by ROM method in 28 articles (Figure 2, Supplementary **Fig. 1-9).** Plasma Aβ42/Aβ40 (0.93, 95%CI: 0.89-0.97), BDNF (0.97, 95%CI: 0.95-0.99) were lower in MCI converters (MCI-c) than MCI nonconverters (MCI-nc). Aβ40 (1.04, 95%CI: 1.02-1.07), T-tau (1.19, 95%CI: 1.11-1.28), P-tau 181 (1.43, 95%CI: 1.31-1.55), P-tau 217 (1.86, 95%CI: 1.53-2.27), NFL (1.23, 95%CI: 1.09-1.39), GFAP (1.65, 95%CI: 1.50-1.81) were higher in MCI-c than MCI-nc. However, the pooled result became unstable after removing Xie 2017 for BDNF (0.95, 95%CI: 0.86-1.05). No differences were found for Aβ42 (1.01, 95%CI: 0.95-1.08) between MCI-c and MCI-nc.

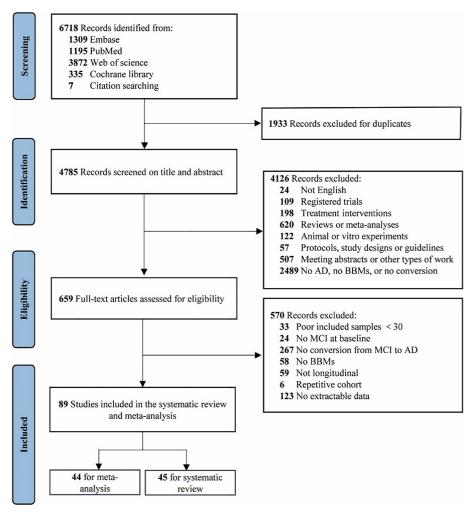


Figure 1. The flow chart of literature selection.

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|--------------------------------------|--------------------|--------------------------|---|---|--------------------------------------|-------------------------------------|---------------------------------------|---|---------------------------------------|---|----------------------|---|------------------------------|
| Blood | | Aβ42 pg/ml | Cystatin C mg/l Aβ42 pg.ml Aβ40 pg/ml Aβ42/Aβ40 | Αβ42 pg/ml Αβ40 pg/ml Αβ42/Αβ40 Αβ pg/ml | Αβ40 ng/L Αβ42 ng/L Αβ42/Αβ40 | Αβ40 ng/L Αβ42 ng/L Αβ42/Αβ40 | Αβ42 pg/ml Αβ40 pg/ml Αβ42/Αβ40 | Αβ42 ng/ml Αβ40 ng/ml Αβ42/Αβ40 | Aβ40 pg/ml Aβ42 pg/ml Aβ42/Aβ40 | APOE | BDNF, log10 ng/mL | BDNF pg/ml | BDNF ng/ml |
| Education Years | Mean (SD) | A A | Ą Z | Ą Z | Υ V | NA | NA | 6.75 (1.62) | 15.82 (2.91) | NA | 15.75 (2.9) | 9.9 (5.7) | Ϋ́ |
| MMSE Mean (SD)/ Meidan | (IOR) | N A | Y Y | N A | 26.95 | 28.77 | 26.2 (2.6) | 26.84 (1.69) | 27.14 (1.75) | 26.86 | 27.11 (2.89) | 26.3 (3) | 26.70 |
| APOE4 Carriers | (%) | A A | 28.57 | A N | 65.60 | 45.31 | 49.60 | 50.52 | ΑĀ | 39.60 | 33.56 | 8 | 51.52 |
| Age Mean (SD)/ Median | (IOR) | N A | 79.79 (4.3) | 75.16 (5.16) | 69.25 (8.05) | 62.36 (6.56) | 75.9 (7.5) | 70.04 (5.63) | 74.65 (7.39) | 76.71 (9.70) | 74.74 (7.32) | 72.2 (6.2) | 71.85 (7.21) |
| Female | | A N | 52.38 | ₹ Z | 59.60 | 48.54 | 55.70 | 62.89 | 34.53 | 61.74 | 35.23 | 75.00 | 46.21 |
| ases | MCI-c | 4 | <u>ო</u> | 37 | 88 | 75 | ΟI | 245 | 145 | e 6 | 161 | 4 | 94 |
| MCI Cases | MCI-nc | 40 | σ | 4 2 | 51 | | 122 | 240 | 162 | 106 | 137 | 42 | 89 |
| MCI Type | | Ū W | MCI | aMCI | aMCI | WC | WC. | aMCI | aMCI | MCI | aMCI | aMCI | aMCI |
| Assay Method (Detection instru- | ments /Antibodies) | ELISA Innogenetics | ELISA 6E10 | ELISA IBL, Gumna Japan | xMAP Innogenetics | xMAP Innogenetics | xMAP Innogenetics | ELISA R162 R165 | xMAP Innogenetics | SOMAscan — | NA | ELISA | ELISA |
| Sample | 246- | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Serum | Serum |
| Follow-up Duration mean/median | (A) | mean 2.5y | mean 4.5y | mean 2y | mean 5.2y range 4-6.8y | mean 2.5y range 1.9-3.9 y | NA | mean 5.2 y range 4.0- 6.8 y | mean 2y | mean 1y | mean 3y | теап 3.9у SD 0.9у | mean 2y |
| AD Criteria | | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | DSM-IIIR NINCDS- ADRDA | NA | NINCDS- ADRDA | DSM-IIIR criteria NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | DSM-IIIR NINCDS- ADRDA |
| MCI | Petersen | 2001 IWG 2004 | CHS cognition study MCI criteria | Petersen 2001 | Petersen 1999 Petersen 2004 | NA | Petersen 1999 IWG 2004 | Petersen 2001 | ADNI Criteria | N A | ADNI Criteria | Petersen 2001 | Petersen 2004 |
| Sample | 2 | Community | Community | Clinical | Clinical | Clinical | Australia Community | Community | Community | Clinical Community | America Community | Clinical | Clinical |
| Region | | Europe | America | Europe | Europe | Europe | Australia | Asia | America | Europe | America | America | Asia |
| Sample Resource | Vienna | Transdanube Aging (VITA) | Pittsburgh Cardiovascular Health Study Cognition Study (CHS-CS) | Ν | Malmö University Hospital | Göteborg MCI study | AIBL | Epidemiology of cognitive Aging (ECA) study | ADNI | EU funded AddNeuroMed (ANM) Alzheimer's Research Trust (ART) | ADNI | Psychogeniatric Clinic of the Institute of Psychiatry, Faculty of Medicine, University of São Paulo | ∀ V |
| Study | | 2008 Blasco | 2008 Lopez | 2009 Cammarata | 2010 Hansson-A | 2010 Hansson-B | 2010 Lui | 2011 Ma | 2011 Toledo | 2014 Kiddle | 2015 Apostolova | 2015 Forlenza | 2015 Wang |

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|--------------|--|---|--|--|-------------------|--|--|---|--|--|--|
| - | Biomarkers | T-tau pg/ml | APOE mg/l | BDNF ng/mL | Αβ40 | Aβ42 pg/ml Aβ42/Aβ40 | Αβ42/Αβ40 | AB42 AB40 T-tau P-tau 181 AB42/AB40 AB42/t-tau AB42/P-tau | T-tau pg/ml Aβ42 pg/ml Aβ40 pg/ml Aβ42/Aβ40 | P-tau 181 pg/ml T-tau pg/ml Aβ42/Aβ40 | Aβ42/Aβ40 |
| Education | Years Mean (SD) | ΑN | A A | 3.38 (1.04) | AA | ΝΑ | 9.20 (4.04) | 11.47 (2.13) | NA | 10 (8-13) | 16.38 (2.5) |
| MMSE | Meidan (IQR) | NA | 26.6 (2.4) | 24.29 | NA | NA | AA | 26.92 (1.58) | NA | 27 (26-28) | 28.72 (1.49) |
| | Carriers (%) | NA | 28 | NA | N | Ν A | 55.20 | 36.40 | N | NA | 48.94 |
| Age | Median Median (IQR) | AA | 67.1 | 71.62 (5.06) | N A | NA | 73.87 (5.11) | 78.55 (2.38) | NA | 73 (69-76) | 68.94 (6.31) |
| | Female | ΑN | 42 | 45.34 | 50.30 | ΝΑ | 51.70 | 36.40 | A N | 43.21 | 48.94 |
| MCI Cases | MCI-c | 20 | 10 | 128 | S S | 265 | 62 | 0 | 0, | 50 Total number for Effect sizes is | |
| MCI | MCI-nc | 103 | 132 | 330 | 121 | 182 | 8 | დ დ | 69 | 80 | 40 |
| | MCI Type | MCI | aMCI | aMCI | aMCI | MCI | aMCI | aMCI | MCI | P-tau 181- ROM: MCI due to AD Effect sizes: MCI | aMCI |
| Assay Method | (Detection instru- ments /Antibodies) | SIMOA — | ELISA - | ELISA - | Immunoassay — | ELISA — | ELISA Araclon Biotech Ltd, Spain | ELISA Immuno-Biological Laboratories Co., Ltd., Japan Invitrogen, Carlsbad, CA, USA Life Technologies Corp., Frederick, MD, USA | IMB | P-tau 181: MSD, Biotinylated-AT270, SULFO-TAG-LRL Aβ42, Aβ40, T-tau: Elecsys | MS |
| 6 | Type | Plasma | Plasma | Serum | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma |
| Follow-up | mean/median (y) | median 3y range 1.1-4.9y | mean 2.3y SD 1.3y | mean 5.08 y SD 0.32y | mean 3y | mean 8.5y | mean 2y | mean 3y | mean 2y | mean 4.9y SD 1.3y | mean By |
| | AD Criteria | NIA-AA 2011 | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | USM-IV | NI-MSO | NIA-AA 2011 NINCDS- ADRDA | NIA-AA 2011 | DSM-5 Aβ+ | NINCDS- ADRDA |
| 2 | Criteria | Petersen 2004 | Petersen 1999 Petersen 2004 NIA-AA | Petersen 1999 | Petersen 2005 | Cognitive impair- ments and CDR | Petersen 2004 | NIA-AA 2011 | NIA-AA 2011 | Petersen 2004 | MMSE score of 24 to 30 CDR-SB score of at least 0.5 |
| | Types | Community | Clinical | Community | Clinical | Community | Olinical | Clinical | Clinical | Clinical | America Community |
| | Region | America | Europe | Asia | Europe America | America | Europe | Asia | Asia | Europe | America |
| | Sample Resource | Mayo Clinic Study of Aging (MCSA) | Amsterdam Dementia Cohort | Mild cognitive impairment and Alzheimer's disease Study in Heibei province (MASHB) | GE067-005 Study | Ginkgo Evaluation of Memory Study (GEMS) | AB255 study | Taipei Veterans General Hospital | National Taiwan University Hospital | Biofinder | ADNI |
| | Study | 2017 Mielke | 2017 van Harten | 2017 Xie | 2018 Westwood | 2019 Lopez | 2019 Pérez- Grijalba | 2019 Chen | 2020 Cheng | 2020 Janelidze | 2020 Shen |

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|--|----------------------------------|------------------------------|--------------------------|--|--|--|--|------------------------|--------------------|---|----------------------------|
| Blood Biomarkers | NFL pg/ml T-tau pg/ml | APOE | NFL pg/ml | GFAP pg/ml | Aβ42/Aβ40: pg/ml p-tau 181 pg/ml NFL pg/ml MS pg/ml P-tau 217 pg/ml | Aβ42/Aβ40: pg/ml p-tau 181 pg/ml NFL pg/ml MS pg/ml MS pg/ml p-tau 217 pg/ml | Aβ42/Aβ40 pg/ml P-tau 181 pg/ml NFL pg/ml | NFL ng/l | P-tau 181 pg/ml | P-tau 217 pg/ml NFL pg/ml | P-tau 181 pg/ml |
| Education Years Mean (SD) | NA | AA | NA | NA | 11.18 | 11.18 | 16.43 | 16.01 (2.82) | AN | Ą | AA |
| MMSE Mean (SD)/ Meidan (IQR) | ΝΑ | A | 27.6 (1.9) | Ą V | 27.21 (1.72) | 27.21 (1.72) | 28.26 (1.74) | 27.93 (1.77) | A A | A | A |
| APOE4 Carriers (%) | A N | ΝΑ | 71.00 | 57.06 | NA | NA | NA | NA | N A | A A | AN A |
| Age Mean (SD)/ Median (IQR) | AN A | ΝΑ | 72.4 (0.7) | 71.61 | 71.36 | 71.36 | 71.51 | 73.48 (7.55) | NA | N A | ΑN |
| Female | ΑN | ΝΑ | 41.91 | 61.29 | 35.50 | 35.50 | 51.20 | 41.80 | N A | ΝΑ | N A |
| MCI Cases | 24 | 400 | 29 | 47 | 145 | 145 | ω | 141 | <u> </u> | 80 | 98 |
| MCI-nc | 88 | 40 | 80 | 79 | 71 | 71 | 98 | 299 | 446 | 80 | 223 |
| MCI Type | MCI due to AD | MCI | aMCI | aMCI | Ξ W | Ξ W | aMCI | aMCI | aMCI | M | aMCI |
| Assay Method (Detection instru- ments /Antibodies) | SIMOA | ×MAP | SIMOA | SIMDA — | Aβ42/Aβ40: Elecsys Aβ42/Aβ40: MS (Aracion Brotech) P-tau 181: MSD (Eli Lilly) NFL: SIMOA P-tau 217: MSD (Blotinylatedil- BA493; SULF GB- TAG-4610-E2) | Aβ42/Aβ40: MS P-tau 181: SIMOA (Quanterix) NFL: SIMOA | Aβ42/Aβ40: MS P-tau 181: SIMOA (Quanterix) NFL: SIMOA | SIMOA (Quanterix) | SIMOA (Quanterix) | P-tau 217 MSD (Bio- tinylated-IBA493, SULFO-TAG-4G10- | SIMOA |
| Sample Type | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma |
| Follow-up Duration mean/median (y) | mean 5.10y SD 2.78y | mean 2.21y | median 4y IQR 4-4.25y | mean 4.7y | mean 4y | mean 4y | mean 4y | mean 4.19y SD 2.33y | mean 3.11y | mean 4y | mean 4y |
| AD Criteria | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS ADRDA | DSM-IIIR NINCDS- ADRDA Aβ+ | VI-WSO | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | VI-MSO | NINCDS- ADRDA |
| MCI Criteria | IWG 2004 | Petersen 2004 IWG 2004 | ADNI Criteria | Petersen 1999 | Petersen 2004 | ADNI Griteria | ADNI Criteria | ADNI Criteria | ADNI Criteria | Petersen 2004 | ADNI Criteria |
| Sample Types | America Community | Olinical | America Community | Clinical | Clinical | America Community | America Community | Community | America Community | Clinical | America Community |
| Region | America | Europe | America | Europe | Europe | America | America | America | America | Europe | America |
| Sample Resource | BU ADC Clinical Core Registry | EMIF-ADMBD- Oxford | ADNI | Memory Clinic at Skåne University Hospital, Malmö | Biofinder | ADNI | ADNI | ADNI | ADNI | Biofinder | ADNI |
| Study | 2020 Sugarman | 2020 Westwood | 2021 Blazhenets | 2021 Cicognola | 2021 Cullen- Biofinder | 2021 Cullen- ADNI | 2021 Cullen- ADNI | 2021 Darmanth'e | 2021 Karikari | 2021 Palmqvist- Biofinder | 2021 Palmqvist- ADNI |

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|-----------------------|--|----------------------------|--|-------------------------|--|---------------------------------------|---|--|--|----------------------------|--|
| Blood | ä | P-tau 181 pg/ml | P-tau 181 pg/ml NFL pg/ml Aβ42 pg/ml Aβ42/Aβ40 pg/ml T-tau pg/ml GFAP pg/ml | P-tau 181 pg/ml | NFL pg/mL [median min-max] APOE ug/ml [median IQB] | Aβ40 pg/ml Aβ42 pg/ml Aβ42/Aβ40 | Αβ42/40 P-tau 181 NFL | NFL pg/ml | P-tau 217 pg/ml NFL pg/ml AB42/AB40 | GFAP, pg/ml | Log-Trans- formed A\$40 pg/ml A\$42 pg/ml A\$42/A\$40 P-tau 181 pg/ml T-tau pg/ml |
| | Years Mean (SD) | NA | 8.97 (4.28) | NA | N A | NA | NA | 9.31 (3.99) | 13.81 | NA | 11.7 |
| MMSE Mean (SD)/ | | NA | 27.21 (1.82) | N | 27.33 | 26.4 (2.5) | NA | 23.28 | NA | NA | 27 (25-29) |
| APOE4 | | NA | 36.40 | N A | 68.97 | 39.60 | 35 | 22.64 | 35 | NA | 25.90 |
| Age Mean (SD)/ | Median (IQR) | NA | 74.47 (5.89) | NA | 64.01 | 77.7 (5.5) | NA | 73.27 (8.35) | 72.29 (8.26) | NA | 71.6 (8.2) |
| | Female | NA | 52.20 | A | 52.45 | 60.40 | 50.00 | 43.40 | 47.27 | NA | 49.80 |
| ases | MCI-c | 77 | 6 | 4 | 28 | 145 | 30 | 0 | 26 | 5 0 | 80 |
| MCI Cases | MCI-nc | 440 | 88 | 584 | 30 | 340 | 30 | 34 | 84 | 2 | 163 |
| | MCI Type | aMCI | aMGI | aMCI | aMCI | MCI | MCI due to AD | MCI due to AD | aMCI | aMCI | aMGI |
| Assay Method | (Detection instru- ments /Antibodies) | SIMOA (Tau12 and AT270) | SIMDA | SIMOA (Quanterix) | NFL: SIMOA (Quanterix) APOE: NA | xMAP Innogenetics | Peoteins: Olink PEA Aβ42, A40: ELISA (Lübeck, Germany) P-tau 181: SIMOA (Quanterix) | SIMOA (Quanterix) | P-tau 217: MSD (Eli Lilly) NFL, AB42, AB40: SIMOA (Quanterix) | GFAP: SIMOA (Quanterix) | SIMOA (Quanterix) |
| Sample | Type | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma | Plasma for others Serum for NFL |
| Follow-up Duration | mean/median (y) | mean 2y | mean 1y | median 4.1y SD 1.34y | mean 2y | mean 3y | mean 5y | mean 2.78y SD 0.63y | mean 3y | mean 3y | median 4.7y range 0.9-8.1y |
| | AD Criteria | NINCDS- ADRDA | DSM V NINCDS- ADRDA | NINCDS- ADRDA | NINCDS- ADRDA | NIA-AA 2011 | NIA-AA2011 | DSM-IV-TR NIA-AA 2011 | NINCDS- ADRDA Aβ⁺ | NINCDS- ADRDA Aβ+ | DSM-IV NINCDS- ADRDA |
| ğ | Criteria | ADNI Criteria | Petersen 1999 | ADNI Criteria | IWG 2004 | Petersen 1999 IWG 2006 | NIA-AA 2011 | NIA-AA 2011 | Petersen 2005 | Petersen 2005 | Petersen 2004 |
| Sample | Types | Community | Clinical | Community | Clinical | Olinical | Clinical | Clinical | Clinical | Clinical | Clinical |
| | Region | America | Europe | America | Europe | Europe | America | Asia | America Europe | America Europe | Asia |
| | Sample Resource | ADNI | ANM | ADNI | University Hospital, Trondheim, Norway | BALTAZAB | Massachusetts Alzheimer's Disease Research Center (MADRC) | Korean Brain Aging Study for the Early Diagnosis and Prediction of AD (KBASE-V) | Multicentter | Multicentter | Shanghai Memory Study (SMS) |
| | Study | 2021 Shen | 2021 Simrén | 2021 Therriault | 2022 Giannisis | 2022 Hanon | 2022 Kivisäkk | 2022 Lee | 2022 Pichet Binette | 2022 Pichet Binette | 2022 Xiao |

| SON | ٥ | _ | œ | 7 | 7 | _ |
|--------------------------------------|--------------------------------------|---|--|------------------------------------|-------------------------------|---|
| Blood Biomarkers | Aß42 pg/ml | P-tau 217 pg/ml P-tau 181 pg/ml P-tau 231 pg/ml | P-tau 181 pg/mL T-tau pg/mL NFL pg/mL GFAP pg/mL | P-tau 181 pg/ml | P-tau 181 pg/mL | GFAP pg/mL NFL pg/mL P-tau 181 pg/mL Aß42/Aß40 T-tau pg/ml |
| Education Years Mean (SD) | 10.58 | NA | NA | NA | ΝΑ | 5.76 (3.76) |
| MMSE Mean (SD)/ Meidan | NA NA | 28 (26-29) | 27.09 | NA | 26.4 (2.5) | NA |
| APOE4 Carriers | 36.67 | 55.60 | NA | NA | 39.80 | 42 |
| Age Mean (SD)/ Median | 69.64 (4.3) | 74 (66-79) | 76.64 (8.11) | NA | 7.7.7 | 66.94 |
| Female | 54.44 | 60.70 | 47.02 | Ν V | 61.40 | 61.00 |
| MCI Cases | | 45 | 47 | 91 | 144 | 90 |
| MCIO | 67 | 06 | 38 | 33 | 332 | 46 |
| MCI Type | aMCI | aMCI | MCI due to AD | aMCI | MGI | MCI due to AD |
| Assay Method (Detection instru- | ×MAP | P-tau 217 MS MSD SIMOA P-tau 181 MS MSD SIMOA Lumipulse P-tau 213 SIMOA | MSD | SIMOA (Quanterix) | SIMOA (Quanterix) | SIMOA (Quanterix) |
| Sample Type | Serum | Plasma | Plasma | Plasma | Plasma | Serum for NFL GFAP Plasma for Aβ42/Aβ40, T-tau, P-tau |
| Follow-up Duration mean/median | (y) mean 2.89y | mean 4.9y SD 2.1y | mean 4y | mean 2.76y SD 0.63y | Range 0.5-3y | mean 5.8y SD 3.4y |
| AD Criteria | NIA-AA 2011 | DSM-IIIR NINODS- ADRDA AR* | NIA-AA2011 | DSM-IV NIA-AA 2011 | DSM IV-TR NINCDS- ADRDA | NIA-AA 2011 |
| MCI Criteria | Petersen 1999 Petersen 2004 | Petersen 2004 | NIA-AA 2011 | NIA-AA 2011 Petersen 2001 | Petersen 2004 | NIA-AA 2011 |
| Sample Types | Clinical | Clinical | Clinical | Clinical | Clinical | Clinical |
| Region | Asia | Europe | America | Asia | Europe | Europe |
| Sample Resource | NBHAD | Memory Clinic at Skåne University Hospital in Malmö | Massachusetts Alzheimer's Disease Research Center (MADRC) | KBASE-V | BALTAZAR | 2023 Silva- Coimbra University Spinola Hospital |
| Study | 2022 Zhao | 2023 Janelidze | 2023 Kivisäkk | 2023 Kwon | 2023 Lehmann | 2023 Silva- Spínola |

Table 1. Characteristics of included studies.

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For variances attributed to MCI types (Fig**ure 2),** the differences between converters and non-converters were significant only in aMCI rather than MCI for Aβ40 and Aβ42/Aβ40. For P-tau 181, the differences were significant not only in a MCI, but also in MCI due to AD and MCI. For NFL, the differences were significant in aMCI but not MCI due to AD. In terms of measuring methods, ELISA assays tested higher Aβ40 (1.069, 95%CI: 1.03-1.11) and lower Aβ42/Aβ40 (0.902, 95%CI: 0.847-0.96) on MCI-c to MCI-nc. SIMOA assays showed lower A\(\beta\)42/A\(\beta\)40 (0.928, 95\(\circ\)CI: 0.896-0.961),

higher T-tau (1.142, 95%CI: 1.049-1.244), P-tau 181 (1.493, 95%CI: 1.36-1.639), P-tau217 (1.746, 95%CI: 1.234-2.47), NFL (1.283, 95%CI: 1.123-1.465) and GFAP (1.675, 95%=1.505-1.865). IMR showed higher T-tau (1.523, 95%CI: 1.245-1.863). MSD showed higher P-tau 181 (1.574, 95%CI: 1.375-1.803), P-tau217 (1.939, 95%CI: 1.233-3.05) and GFAP (1.108, 95%CI: 0.875-1.404). MS showed higher P-tau 181 (1.162, 95%CI: 1.038-1.3) and P-tau 217 (1.894, 95%CI: 1.477-2.43). As outlined above, the pooled ROMs vary depending MCI types and assays, which should be underscored in future studies.

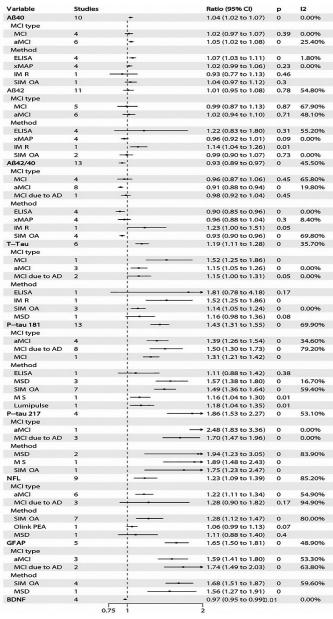


Figure 2. MCI converters to MCI non-converters ratio of blood biomarkers.

3.2.2. Relative risks of blood biomarkers

Relative risks (RR) of plasma Aβ42, Aβ42/Aβ40, P-tau, NFL, and APOE were pooled as continuous or categorical variables in 22 articles (Figure 3, Supplementary Fig. 10-13). P-tau (RR=2.50, 95%CI: 2.04-3.06) as a continuous variable, Aβ42/ Aβ40 (RR=1.28, 95%CI: 1.01-1.61) as a categorical variable can predict the conversion from MCI to AD. Aβ42/Aβ40 (RR=0.97, 95%CI: 0.84-1.13), NFL (RR=1.20, 95%CI: 0.78-1.85) as continuous variables, Aβ42 (RR=1.01, 95%CI: 0.74-1.39) as

categorical variable couldn't predict the status conversion of MCI. Besides, APOE (RR=1.03, 95%CI: 0.89-1.19) couldn't predict MCI conversion either.

When it comes to specific MCI types, continuous P-tau could predict the conversion of either aMCI (RR=2.152, 95%CI: 1.661-2.788) or MCI (RR=3.188, 95%CI: 2.29-4.439). However, categorical Aβ42/Aβ40 could neither predict the conversion of sole aMCI group (RR=1.37, 95%CI: 0.944-1.987) nor the conversion of the sole MCI group (RR=1.222, 95%CI: 0.912-1.636).

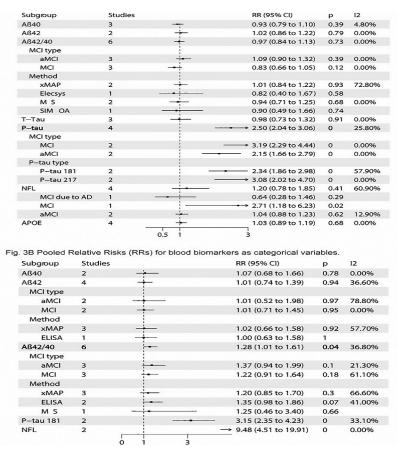


Figure 3. Pooled relative risks (RRs) for blood biomarkers as continuous or categorical variables.

3.2.3. Diagnostic performance of blood biomarkers

Diagnostic indexes of Aβ42/Aβ40, P-tau 181, P-tau 217, NFL, and GFAP were pooled in 12 articles (Figure 4, Supplementary Fig. 14-22). The HS-ROC curves of the pooled results demonstrated that the diagnostic efficacy of A β 42/A β 40 (P=0.102), P-tau 181 (P=0.788), P-tau 217 (P=0.978), NFL

(P=0.579) was not impacted by threshold values. However, the diagnostic efficacy of GFAP was influenced by the threshold effect (P=0.000). For Aβ42/Aβ40, the pooled sensitivity and specificity were 86% (95% CI: 0.80-0.91) and 49% (95% CI: 0.25-0.73), respectively. The pooled PLR and NLR were 1.7 and 0.28, respectively. The pooled DOR was 6 (95% CI: 3-13), with an AUC of 0.84 (95% CI: 0.81-0.87). For P-tau 181, the pooled sensitivity

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and specificity were 81% (95%CI: 0.74-0.87) and 78% (95%CI: 0.70-0.84), respectively. The pooled PLR and NLR were 3.6 and 0.24, respectively. The pooled DOR was 15 (95% CI: 10-23), with an AUC of 0.86 (95% CI: 0.83-0.89). For P-tau 217, the pooled sensitivity and specificity were 91% (95%CI: 0.85-0.94) and 81% (95%CI: 0.76-0.86), respectively. The pooled PLR and NLR were 4.9 and 0.12, respectively. The pooled DOR was 42 (95% CI: 21-81), with an AUC of 0.93 (95% CI: 0.91-0.95). For NFL, the pooled sensitivity and specificity were 80% (95%CI: 0.69-0.87) and 61% (95%CI: 0.46-0.75), respectively. The pooled PLR and NLR were 2.1 and 0.33, respectively. The pooled DOR was 6 (95% CI: 3-13), with an AUC of 0.80 (95% CI: 0.76-0.83). For GFAP, the pooled sensitivity and specificity were 71% (95%CI: 0.63-0.79) and 86% (95%CI: 0.81-0.91), respectively. The pooled PLR and NLR were 4.33 and 0.39, respectively. The pooled DOR was 12.71 (95% CI: 7.71-22.54), with an AUC of 0.84 (95% CI: 0.78-0.90). The Deek's funnel plot asymmetry test results did not show publication bias for the above five biomarkers (Aβ42/Aβ40: P=0.860; P-tau 181: P=0.893; P-tau 217: P=0.470; NFL: P=0.426; GFAP: P=0.801). In subgroup analysis by MCI type, Aβ42/Aβ40 had higher sensitivity and specificity in aMCI than MCI, while NFL had higher sensitivity and specificity in MCI than aMCI (Supplementary material 6).

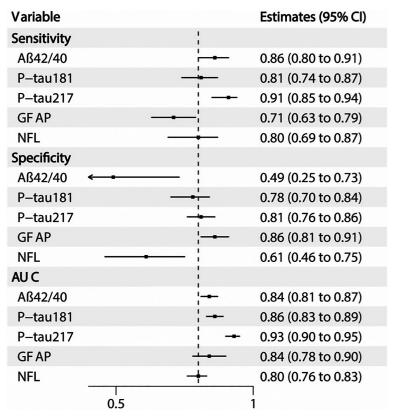


Figure 4. Pooled sensitivity, specificity, and AUC for blood biomarkers in predicting MCI conversion.

3.2. Subgroup analysis, meta-regression, sensitivity analysis, and publication bias

Heterogeneity existed among Aβ42, Aβ42/Aβ40, P-tau181, P-tau217, NFL and GFAP in ROM, NFL in continuous RR, Aβ42/Aβ40, P-tau181 and NFL in diagnostic analyses. Subgroup analyses (Supplementary Fig. 23-59) showed that baseline MMSE, female percentage, APOE carries and region might be the sources of heterogeneity for P-tau 217 in ROM, baseline MMSE, age and follow-up duration for GFAP in ROM, study type, region and MCI type for RR of NFL as continuous variables. For diagnostic results, APOE $\varepsilon 4$ carriers and study type for A β 42/A β 40, study type for P-tau 181, and follow-up duration for NFL are potential factors

bringing about heterogeneity, as demonstrated by meta-regression results (Supplementary material 5). Sources of heterogeneity have not been found for Aβ42, Aβ42/Aβ40, P-tau 181, and NFL in ROM. However, the stability of the pooled ratios for P-tau 181 and NFL was stable, whereas unstable for Aβ42 and Aβ42/Aβ40 after sensitivity analyses (Supple**mentary Fig. 60-68).** The heterogeneity of Aβ42, $A\beta42/A\beta40$ (I²=54.8% P=0.015; I²=45.5% P=0.03) has decreased but was still significant after removing an article (Ma 2011). In a new round of subgroup analysis, baseline MMSE and female percentage for Aβ42, baseline age and female percentage for Aβ42/Aβ40 contributed to the heterogeneity. Meanwhile, the pooled result of Aβ42 and Aβ42/Aβ40 became stable after sensitivity analyses. In the end, trim-and-fill method (Supplementary Fig. 69-73) indicated that the pooled RRs of continuous NFL and categorical Aβ42/Aβ40 were unstable. Therefore, the pooled results should be interpreted with caution. Publication bias (Supplementary Fig. 74-**93)** was found among P-tau181 (P=0.041) in ROM. Nevertheless, the effect size of P-tau181 was not influenced after implementing the trim-and-fill method.

3.3. Qualitative review

A total of 80 biomarkers without sufficient data to carry out meta-analysis were summarized here. Under the ATNIVS frame (Table 2), categorical P-tau 181 and NFL can predict the risk of MCI conversion, while continuous Aβ42, Aβ40, and categorical Aβ40 could not (**Figure 3**). However, caution is needed in case of over-interpretation, given the limited number of included studies.

For other blood biomarkers, continuous T-tau couldn't predict MCI conversion (Figure 3A). Higher coated platelet levels, beta-secretase 1 (BACE1) activity, cortisol, APOA-II, soluble TREM2 (sTREM2), sTNFR1, CCL23, IL-17A, mid regional proadrenomedullin (MR-ProADM), midregional proatrial natriuretic peptide (MR-ProANP), nonceruloplasmin copper, miR-146a, miR-181a, lower APP ratio (APPr), intracellular calcium-independent PLA2 (iPLA2), complement C3 (CC3), complement factor I (CFI), CCL11, not dissociated Aβ42 autoantibodies, HSV-1 specific antibodies, epidermal growth factor (EGF), neural growth factor (NGF), TSH, 24-hydroxycholesteryl esters (24OH-CE), miR206, ceruloplasmin, selenoprotein P and caffeine were related with increased risk of MCI conversion. Nevertheless, the predictive performance of some BBMs such as APOA-I, clusterin, total cholesterol, homocysteine, cystatin C, CRP, complement factor H (CFH), transthyretin, pancreatic prohormone, uric acid, MDA, Ficolin-2, serum calcium, miR132 and alpha-2-macrogloblin (A2M) remained underdetermined due to contradictory results from insufficient research. Notably, neuronal-derived exosome (NDE) is an emerging candidate in the diagnosis and prognosis of AD. Lower levels of Aβ1–42 in plasma NDEs predict MCI conversion with an AUC of 0.84. Besides, plasma NDE levels of P-S396-tau, repressor element 1-silencing transcription factor (REST), and neurogranin (NRGN) take on remarkable discrimination of MCI-converters and MCI-nonconverters.

Furthermore, proteomics and metabolomics uncover the potential values of other blood biomarkers. For proteomics, inflammation/chemotaxis (IL-8, CSF-1, CCL23, CX3CL1, CXCL and TNFRSF12A),

| Biomarker category | Higher | Lower | Undetermined | | | | | |
|---|----------------------------------|---------------------------|----------------|--|--|--|--|--|
| Core biomarkers | | | | | | | | |
| A (Aβ proteinopathy) | BACE1 activity | Appr | clusterin, A2M | | | | | |
| T (AD tau proteinopathy) | categorical P-tau 181 | _ | _ | | | | | |
| Non-specific biomarkers of tissue reaction involved in AD pathophysiology | | | | | | | | |
| N (injury, dysfunction, or degeneration of neuropil) | categorical NFL | _ | _ | | | | | |
| I (inflammation) | sTREM2, sTNFR1, CCL23, IL-17A | iPLA2, CC3, CFI, CCL11 | CRP, CFH | | | | | |
| Biomarkers of non-AD co-pathology | | | | | | | | |
| V vascular brain injury | MR-ProADM, MR-ProANP | _ | _ | | | | | |
| S α-syn | _ | _ | _ | | | | | |

Table 2. Blood biomarkers included in qualitative review.

extracellular matrix remodeling (TIMP-4 and MMP-3), endothelial injury (VEGF-A and NOS3), lipid metabolism (PHOSPHO1) and insulin-like growth factor signaling regulation (IGFBP2), proprotein convertase subtilisin/kexin type 7 (PCSK7), ephrin receptor tyrosine kinase A2 (EFNA2), AP-1 complex subunit gamma-like 2 (AP-1), and collagen alpha1 (XV) chain (COL15A1) were reported to be related to MCI conversion. For metabolomics, 2,4-dihydroxybutanoic acid (MC1), Unidentified carboxylic acid (MC2), PC aa C38:4, PC ae C36:2, PC ae C40:3, PC ae C42:4, PC ae C44:4, PC ae C44:4, SM C16:0, SM C18:1, SM (OH) C14:1, SM C20:2, polyamine and l-arginine metabolism were implicated in the conversion. Collectively, plentiful blood biomarkers hold the potential to predict MCI conversion. However, more validation studies are warranted.

4. DISCUSSION

In the present study, we conducted a comprehensive systematic review and meta-analysis aiming to evaluate the predictive effects of blood biomarkers in the conversion from MCI to AD. Our study first demonstrated that baseline levels of Aβ40, Aβ42/ Aβ40, T-tau, P-tau 181, P-tau 217, NFL, and GFAP were discrepant among MCI converters from MCI non-converters by analyzing pooled ratio of means. Secondly, we found baseline continuous P-tau, categorical Aβ42/Aβ40, could predict the future conversion risk of MCI by pooling RRs. Thirdly, diagnostic meta-analyses exhibited that Aβ42/Aβ40, P-tau 181, NFL, and GFAP exhibited superior value, while P-tau 217 manifested remarkable value in predicting MCI conversion to AD.

Plasma Aβ42/Aβ40 is associated with CSF $A\beta42/A\beta40$ and $A\beta$ -PET and is more valuable in predicting conversion to MCI or AD in cognitively unimpaired (CU) people than A β 42 or A β 40 alone. Alternatively, plasma Aβ42/Aβ40 has been completely altered in the pre-symptomatic stage of AD, rendering it possible to accurately identify Aβ pathology in CU people (Palmqvist et al., 2019). Our pooled results suggested lower Aβ42/Aβ40 and higher Aβ40 in MCI-c compared to MCI-nc, which were discordant with a former meta-analysis (Qu et al., 2021). Possible explanations were that we included more recent studies and removed one study that brought significant heterogeneity (Fei, Jianghua, Rujuan, Wei, & Qian, 2011). Nevertheless, aligning with Li et al., (Li, Ma, Tan, & Yu, 2022),

we found that categorical Aβ42/Aβ40 could predict future MCI conversion. In diagnostic performances, the DOR of $A\beta42/A\beta40$ is 6, which is lower than P-tau 217, P-tau 181, GFAP, suggesting that Aβ42/ Aβ40 might not be that sensitive to the early progression of Alzheimer's continuum, and it might have reached a plateau at preclinical AD (Yakoub et al., 2023) Besides, its specificity requires improving. Moreover, the predictive performances of plasma Aβ42/Aβ40 were lower than CSF Aβ42/Aβ40 (Li et al., 2022). The possible explanations are as follows. Firstly, the stickiness of Aβ hinders its flow into the bloodstream, and its transportation mechanisms remain unclear. Secondly, detecting soluble Aβ in plasma is challenging due to blood dilution, and its level decreases as AD progresses. Thirdly, derived from the APP protein, Aβ is not only confined to the CNS and but also serves physiological functions, including antimicrobial roles, rendering peripheral Aß production and clearance complex. At last, the amphipathic structure of AB causes it to bind to various proteins and blood cells (Huang, Wang, & Guo, 2022).

As well-known, the advent of blood P-tau seems to have altered the landscape of AD diagnosis and prognosis since it increases with Aß accumulation and clinical severity in patients rather than in individuals with non-AD induced cognitive impairment (Karikari et al., 2022). Moreover, even in people without Aβ pathologic deposits at baseline, P-tau 231 and P-tau 217 were in close relation to a longitudinal increase in Aβ-PET uptake. Chronologically, P-tau181 continued to rise as CDR scores went from 0 to 3, but did not reach abnormal levels until 6.5 and 5.7 years after abnormalities CSF Aβ and Aβ-PET, respectively (Moscoso et al., 2021), whereas alterations of plasma P-tau 231 and P-tau217 occurred before the appearance of pathological Aß plaques, thus reflecting early brain Aß changes earlier. Thus, plasma P-tau 231 and P-tau 217 are earlier biomarkers indicating AB changes than P-tau 181. Particularly, P-tau231, may be the earliest abnormal blood p-tau of AD (Ashton et al., 2022; Milà-Alomà et al., 2022). However, in our current review, only one study investigated the predictive performance of P-tau 231 in MCI conversion (Janelidze, Bali, et al., 2023) (sensitivity: 0.87; specificity: 0.69; AUC:75%). Besides, it might have also reached a plateau at preclinical AD. Therefore, further studies of P-231 are warranted (Yakoub et al., 2023). Interestingly, plasma P-tau181 and P-tau217 have been consecutively

reported to accurately predict cognitive deterioration and conversion to AD dementia in the next 2-6 years in MCI patients (Hansson et al., 2022). In consistent with previous studies, our pooled results also attested to the predictive ability of P-tau 181 and P-tau 217. Given that P-tau 217 has the largest relative increase (250%-600%) in AD compared to other non-AD neurodegenerative diseases (Thijssen et al., 2021), it might be the most sensitive blood biomarker to detect MCI conversion, which was further proved by its pooled sensitivity, specificity and DOR. Nonetheless, although plasma P-tau has gained momentum in the timely diagnosis of AD, it can elevate various comorbidities (such as chronic kidney disease, hypertension, etc.), giving rise to false positives (Mielke et al., 2022). Fortunately, a recent study indicated that applying P-tau/Ttau could mitigate the impacts on kidney function (Janelidze, Barthélemy, He, Bateman, & Hansson, 2023).

As for T-tau, the overlap of T-tau between diagnostic groups is considerable, which has limited its diagnostic value. The correlation between T-tau levels in blood and CSF was also poor, indicating that the majority of T-tau in the blood is of peripheral origin. In contrast, the central origin is relatively small (about 20%), making it impossible to detect ongoing neurodegeneration. Nevertheless, high baseline levels of T-tau often predicted faster cognitive decline (Karikari et al., 2022), which was further corroborated by our finding that MCI converters have higher baseline T-tau than MCI non-converters.

Unlike other blood biomarkers of AD, serum GFAP (a marker of reactive astrocyte hyperplasia) is far superior to CSF GFAP in determining the pathological exacerbation of brain Aß (Benedet et al., 2021) since periodic freeze-thaw has a significant effect on the concentration of CSF GFAP rather than serum GFAP (Simrén, Weninger, et al., 2022). Serum GFAP begins to rise in preclinical AD and is correlated with AD incidence over 10 years before diagnosis (9-17 years). P-tau181 and NfL were related to moderate AD risk (up to 9 years), suggesting that GFAP might be an earlier biomarker before P-tau181 and NFL for AD (Stocker et al., 2023), which has also been confirmed by recent studies (Milà-Alomà et al., 2022). Besides, serum GFAP can also distinguish AD from frontotemporal dementia, and predict the cognitive decline of CU (Hansson et al., 2022) and the transition of MCI to dementia (Oeckl et al., 2022). Therefore, plasma GFAP seems to be a very promising biomarker. The present findings also disclosed higher GFAP in MCI converters and higher DOR compared to NFL and $A\beta 42/A\beta 40$.

As a marker of nerve axon injury, NFL can be detected simultaneously in CSF and plasma (serum), and is currently the most promising marker of neurodegeneration. In cognitively normal people, plasma NFL increases markedly with age and becomes more pronounced after age 65 (Simrén, Andreasson, et al., 2022). Although NFL is weak as a diagnostic biomarker relative to P-tau 181 or P-tau 231, above-threshold NFL levels predict faster cognitive decline (Smirnov et al., 2022). The pooled ROMs indicated higher NFL in MCI converters, yet pooled RRs suggested no predictive effects of NFL, which contradicted Li et al., (Li et al., 2022). The reasons for these discrepancies might be that we concentrated mainly on MCI conversion, while Li et al. focused on the whole AD continuum. Ultimately, it's worth noting that age has an increasing impact on serum NFL levels, while other factors such as kidney function and blood volume also have some effects (Koini et al., 2021) among people over 60 years old.

Generally, given that MCI is multifactorial in etiology and heterogeneous in clinical presentation, the predictive value of single biomarkers tends to be limited. Therefore, the combination of different BBMs or of BBMs with biomarkers from other easily accessible categories might contain giant potential. It's reported that amnestic MCI patients with hAT (lower plasma Aβ42/Aβ40 and higher p-tau181) had 4.83 times the risk for AD conversion (HR=4.83, 95% CI 2.37–9.86, P<0.001) compared to patients with lAT (higher plasma Aβ42/Aβ40 and lower p-tau181) (Xiao et al., 2022). Plasma P-tau 217 combined with cognitive tests and APOE &4 status forecasted the conversion to AD in MCI patients with high accuracy (AUC: 0.89, 95%CI: 0.83-0.94) (Janelidze et al., 2020; Palmqvist et al., 2022).

Limitations in our meta-analysis should be acknowledged. First, sources of heterogeneity still couldn't be found after meta-regression and subgroup analysis for P-tau 181 and NFL in ROM. Other variables (e.g., education years, adjustments, and medications) should also be considered in future studies. In addition, data extraction might bring some heterogeneity. Specifically, some studies reported log-transformed value; some reported median (IQR), while there are a handful of studies whose data couldn't be directly extracted from articles

nor attained from the authors, which could only be manually acquired from figures. Secondly, current studies were relatively few and lacked detailed information such as capture antibodies. Therefore, the results of subgroup analysis should be interpreted with caution. Meanwhile, the predictive performances of some biomarkers might be underestimated due to insufficient research. More studies were warranted to validate our results. Last but not least, included studies are primarily conducted by some cohorts utilizing retrospective samples in specialized centers. Prospective validation and inclusion of particular types of MCI are necessary for future research.

5. CONCLUSION

This meta-analysis has discussed the predictive role of blood biomarkers in the conversion from MCI to AD, which will facilitate the clinical implementation of blood biomarkers soon. Of note, the predictive effects may vary with MCI types, as well as measuring methods. Therefore, more prospective cohorts based on particular MCI and high-sensitivity assays are needed to validate our current results.

Acknowledgments

None.

Conflict of Interest Disclosures

The authors declare no conflict of interest.

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Data Availability Statement

The data that support the findings of this study are available on request from the corresponding author upon reasonable request.

LIST OF ABBREVIATIONS

| Abbreviation | Full name |
|--------------|---|
| 240H-CE | 24-Hydroxycholesteryl Esters |
| A2M | alpha-2-macrogloblin |
| Αβ | amyloid-β |
| AD | Alzheimer's disease |
| APOA-II | apolipoprotein-ll |
| APOE | apolipoprotein E |
| APP | amyloid precursor protein |
| BACE1 | beta-site amyloid precursor protein cleaving enzyme 1 |
| BBM | blood biomarkers |
| BDNF | brain-derived neurotrophic factor |
| CXCL | chemokine (C-X-C motif) ligand |
| EGF | epidermal growth factor |
| GFAP | glial fibrillary acidic protein |
| iPLA2 | intracellular calcium-independent PLA2 |
| IGFBP2 | insulin-like growth factor signaling regulation |
| MR-ProADM | midregional proadrenomedullin |
| MR-ProANP | midregional proatrial natriuretic peptide |
| NDE | neuronal-derived exosome |
| NFL | neurofilament light |
| NGF | neural growth factor |

| Abbreviation | Full name |
|--------------|--|
| NRGN | neurogranin |
| p-tau | phosphorylated tau |
| PCSK7 | proprotein convertase subtilisin/kexin type 7 |
| REST | repressor element 1-silencing transcription factor |
| sTNFR1 | soluble tumor necrosis factor α receptor 1 |
| sTREM2 | soluble form of triggering receptor expressed on myeloid cells 2 |
| t-tau | total tau |
| TNFRSF12A | TNF receptor superfamily member 12A |

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SUPPLEMENTARY MATERIALS

Supplementary material 1: PRISMA checklist **Supplementary material 2:** Search strategy

Supplementary material 3: References for included 89 studies

Supplementary material 4: Included studies for pooled ROMs, RRs, and diagnostic indexes

Supplementary material 5: NOS score for studies included in meta-analysis

Supplementary material 6: Meta regression



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